

The c.3140-26A>G Variant of the *CFTR* Gene in Homozygous State Causes Mild Cystic Fibrosis – Overview of Longitudinal Clinical Data of the Patient Managed in our CF Center and Review of the Literature

Ana Kotnik Pirš,^{1,2*} Uroš Krivec,¹ Katarina Trebušak Podkrajšek,^{3,4}

¹Unit for Pulmonary Diseases, University Children's Hospital, University Medical Centre Ljubljana, Bohoričeva 20, 1000 Ljubljana, Slovenia

²Department for Pediatrics, Faculty of Medicine, University of Ljubljana, Bohoričeva 20, 1000 Ljubljana, Slovenia

³Institute of Biochemistry, Faculty of Medicine, University of Ljubljana, Vrazov trg 2, 1000 Ljubljana, Slovenia

⁴Clinical Institute for Special Laboratory Diagnostics, University Children's Hospital, University Medical Centre Ljubljana, Vrazov trg 1, 1000 Ljubljana, Slovenia

*Corresponding author: Ana Kotnik Pirš, MD, PhD. E-mail: ana.kotnikpirs@kclj.si. Unit for Pulmonary Diseases, University Children's Hospital, University Clinical Centre Ljubljana and Department for Pediatrics, Faculty of Medicine, University of Ljubljana, Bohoričeva 20, 1000 Ljubljana, Slovenia. Tel.: +386 1 522 9257; Fax: + 386 1 522 4073.

Abstract

There are over 70.000 patients with cystic fibrosis (CF) in the world and numerous sequence variations in the *CFTR* gene have been reported but the clinical significance of all of them is still not known. There are currently 195 patients with the c.3140-26A>G (legacy name 3272-26A>G) variant in the *CFTR* gene listed in the European Cystic Fibrosis Society Patient Registry (ECFSPR) and only 4 are homozygous. We present longitudinal clinical data of one of these patients who is managed in our CF Center at the University Children's Hospital in Ljubljana and compare it with the patient data from the ECFSPR and the CFTR2 database in which additional 3 homozygous patients are described.

Moreover, the effect of the detected variant in the described patient was evaluated on the RNA level in nasal epithelial cells. The variant was shown to result in aberrant splicing introducing a frameshift and a premature termination codon while normal transcript was not detected. Alternative spliced mutant transcripts in other tissues or the presence of

spliceosome-mediated RNA trans-splicing could explain the mild clinical presentation of patients with this variant in homozygous state.

Key words: cystic fibrosis, *CFTR* gene, c.3140-26A>G variant, homozygous, aberrant splicing, longitudinal patient data

1. Introduction

1.1. Cystic fibrosis

Cystic fibrosis (CF) is one of the most common inherited diseases in Caucasians. The incidence of the disease differs between populations with the highest incidence in Ireland 1:1.800 and the lowest in Finland with 1:20.000.¹ The incidence of CF in Slovenia is 1:4.500.² CF affects many organs. The lungs, gastrointestinal system, pancreas, liver, nose and sweat glands are most commonly involved. In 1989 the *CFTR* gene was discovered and variants of this gene were shown to be causative of CF.³ In 2016 De Boeck and Amaral⁴ updated the 1995 Zielinski and Tsui⁵ classification of *CFTR* variants from 5 classes to 7 according to the mechanism by which they disrupt the synthesis, trafficking and function of the CFTR protein. CF is diagnosed on the basis of clinical presentation, sweat testing and genetic confirmation.⁶ To date there are over 44.000 sequence variants listed in the Ensembl genome browser 98 (<https://www.ensembl.org/index.html>; accessed in October 2019) in the *CFTR* gene, but not all of them are disease causing. In October 2019, more than 1800 CF causing variants located in the coding region, splice sites and the regulatory regions of the *CFTR* gene have been reported in the professional version of the Human Gene Mutation Database.⁷

Incredible progress has been made in the past 10 years in the treatment of CF. Regular therapy that improves the symptoms which are a consequence of CFTR dysfunction has been a cornerstone of CF therapy since the implementation of pancreatic enzyme replacement therapy (PERT) shortly after the first description of CF by Dorothy Hansine Andersen in 1938.⁸ New medications that influence the CFTR directly have been developed and implemented into regular therapy in 2014. New drugs named CFTR modulators work in two ways – as potentiators they enhance the function of the CFTR on the apical cell membrane in patients with milder *CFTR* variants in which the number or the function of the CFTR is affected. The second class are CFTR correctors – they influence the translation, folding and transport of the CFTR to the apical membrane in patients with severe variants in which there

is only a small number of dysfunctional CFTR on the apical membrane or none at all. Potentiators (ivacaftor) are beneficial in patients with splicing variants as are the patients presented in our review, while a combination of potentiators and correctors, such as the recently approved triple combination of elexacaftor/ivacaftor/tezacaftor, is needed in patients with the classic severe *CFTR* variants such as the common p.Phe508del variant.⁹

Even with all the progress that has been made, most leading authorities in the field of CF believe that only close follow-up of patients can show the effect of a particular variant on a particular patient. It is widely accepted that two patients with the same disease causing variant can have an entirely different clinical progress of CF.^{10, 11}

1.2. The c.3140-26A>G *CFTR* variant

According to the CFTR1 and CFTR2 mutation databases and the so far published literature, the c.3140-26A>G variant (legacy name 3272-26A>G) is considered to be a mild variant of the *CFTR* gene.^{12, 13} Based on the combination of clinical and functional evaluation it is listed as a disease causing variant. According to CFTR1 and CFTR2, this variant does not need functional testing.¹²⁻¹⁴ Most of the patients in the European Cystic Fibrosis Society Patient Registry (ECFS PR) and the CFTR2 database are compound heterozygotes, only 7 patients altogether are homozygotes. Therefore, it is believed that the variant is clinically important especially when in combination with another disease causing variant. Patients with this variant are likely to be pancreatic sufficient.¹³ According to the published literature patients have mild signs of pulmonary disease. Some authors report that nasal polyps and chronic rhinosinusitis are more common.¹⁵⁻¹⁷ After a thorough search through the National center for biotechnology information (NCBI) databases on this variant up to October 2019 no comprehensive clinical reports on homozygous patients with this variant have been published so far. We describe 12 year longitudinal clinical data on a homozygous patient with this variant managed in our CF Center.

2. Methods

All existing medical records were analyzed and clinical examinations were performed as recommended by the ECFS DNWG.⁶ Evaluations were performed at the University Children's Hospital, University Medical Centre Ljubljana, Slovenia, where written informed consent was

obtained from the parents prior to the study and Declaration of Helsinki protocols were followed.

Genomic DNA was isolated from blood leucocytes using the FlexiGene DNA Kit 250 (Qiagen, Germany) according to an established laboratory protocol. Initial mutation screening with Oligonucleotide Ligation Cystic Fibrosis Assay (Abbott, Germany) did not detect any of the 32 analyzed variants. Therefore, the *CFTR* coding region together with exon-intron boundaries were PCR amplified using in-house designed primers (Table 1) and sequenced with the BigDye Terminator v.3.1 Cycle Sequencing Kit and the 3500 Genetic Analyzer capillary electrophoresis system (Life Technologies, USA). Identified sequence variants were confirmed by sequencing of the additional independent PCR amplicon.

To demonstrate the effect of the identified c.3140-26A>G variant *in vivo*, total RNA of the patient, her mother and two unrelated healthy control subjects was purified from nasal epithelial cells using QIAzol Lysis Reagent and Qiagen RNeasyMini Kit (Qiagen, Germany) according to the manufacturer's instructions. Reverse transcription of the total RNA was performed with random hexamers (Applied Biosystem, Foster City, CA, USA) and SuperScript II Reverse Transcriptase (Invitrogen, USA). *CFTR* cDNA was amplified with primer pair aligning exon 16 (forward primer) and exon 17B (reverse primer) (Table 1) and sequenced. Additionally, control amplification of the *TP53* gene was performed.

Table 1. Primers used for the sequencing of the *CFTR* gene coding region and cDNA.

Exon nr.	Exon nr. (legacy)	sequence 5'-3'	Amplicon length (bp)	Cycling conditions
1	1	cagcactcggcttttaacct catacacacgccctcctctt	336	Annealing temperature: 59°C;
2	2	ttccatatgccagaaaagttga gccaccatactggctccta	328	
3	3	ccttgatatacttggtgaatcaa ttggagttggattcatcctt	332	
4	4	aaactgtctcccactgttgct ttaattcagcatttatcccttacttg	543	
5	5	gcctagatgctgggaaataaaac ttactattatctgacccaggaaaactc	411	
6	6a	ttgttagtttctaggggtggaaga gcagtcctgggtttactaaagtgg	305	
7	6b	gggatagagatagcatatggaatgag acaaacatcaaataatgaggtggaag	449	
8	7	catgctcagatcttccattcc tttctatcttttcgcacatttt	431	
9	8	cattagtggttaattcagggttg ggatgaaatccatattcacaaga	497	
10	9	ggccatgtgcttttcaaact ctccaaaataccttcagcac	389	

11	10	caagtgaatcctgagcgtga tccattcacagtagcttacccata	387	
12	11	tgaatttgtaaaatggacctatgga caattccagaacagaatataaagca	533	
13	12	gtaatgcatgtagtgaactgtttaagg tttagcatgaggcgggtgag	398	
16	14b	gcatgggagggaatagggtgaa gcctgtggaggagctagga	305	
17	15	ccatttacatgtattggaaattcag acaaaaccacaggccctatt	398	
18	16	aatgcgtctactgtgatccaaac gacaggactcaaccctcaatc	288	
19	17a	caatgtgaaaatgtttactaccaac ttgggaacccagagaaacct	542	
20	17b	caaagaatggcaccagtggtg aaacaatggaaattcaaagaaatca	387	
21	18	tgatatgtgccctaggagaagtg cagtgaacctcaattatctgtaatg	329	
22	19	aaagcccgcacaaataaccaa tcaggctactgggattcattac	443	
23	20	ttccactggtgacaggataaaa aaaaagacagcaatgcatacaaaa	399	
24	21	tgatggtaagtacatgggtgtttc aaaatcatttcagttagggttaggt	411	
27	24	ccagtttctgtccctgtctct gtgtcctcaattccccttacc	439	
14	13	gagagaccccaggagataatg tgcattctgtgggggtgaaa	1066	Annealing temperature: 59°C; elongation 1'
15	14a	accacaatggtggcatgaa ctccccactcagtcctctaaaa	791	
25, 26	22, 23	gaatgtcaactgcttgagtgtttt tgagtaaagctggatggctgt	1081	
cDNA: 18, 19, 20	cDNA: 16, 17a, 17b	ccaaacctcacagcaactca tggcctaaagtgcaggataatac	308	Annealing temperature: 58°C

3. Results

3.1. Clinical characteristics

In 2007 a 12-year-old girl with unrelated parents and no family history of chronic disease was referred to our pediatric pulmonologist clinic because of persistent dry cough. The girl was born at term as the second child after a second uneventful pregnancy and birth. She was gaining weight and height well. At the age of 4 years she had a viral pneumonia without any complications. From the age of 9 she had nasal discharge, tearing and sneezing especially in the spring. From the age of 11 she was coughing almost persistently particularly when in contact with dust mite, after exertion and while laughing. She had no history of diarrhea, bulky, fatty feces, stomach discomfort or distention.

At the time of her first visit, the girl had a healthy appearance. Her weight, height and body mass index were on the 75th, 99th and 34th percentile for age and gender respectively. The

clinical examination was normal beside a partially blocked nose. On spirometry the forced vital capacity (FVC) was 80% of predicted values for age, height and gender, the forced expiratory volume in one second (FEV1) 72% and the Tiffenau index 89. Asthma was excluded with a negative Metacholine Challenge Test. She had normal levels of exhaled and nasal nitric oxide (FeNO). Allergies were excluded with skin prick testing and blood IgE evaluation and her chest X-ray was normal.

She returned to our outpatient clinic after 3 years because of persistent and worsening cough. She had no other signs or symptoms of CF. Sweat testing was performed using the Gibson-Cooke method. The sweat chloride levels were elevated on two separate examinations: 56 and 57.7 mmol/l – the high end of intermediate levels. In concordance with the European recommendations for molecular genetic diagnosis of CF and CFTR-related disorders¹⁷ molecular diagnostic testing was issued confirming the diagnosis of CF with identification of the c.3140-26A>G variant in homozygous state.

For the last 12 years, since the diagnosis was confirmed, the patient has been managed in our center for children and adolescents with CF according to the standards of care.¹⁸ She is pancreatic sufficient with normal fecal elastase levels, has normal liver function tests and abdominal ultrasound, normal levels of A, D, E, K vitamins without supplementation, glucose metabolism and bone density. She has chronic rhinitis, but no nasal polyps. Her chest x-ray is normal therefore a chest CT has not been done yet. Her FEV1 is currently 70% predicted, with a yearly decline of 0.6% despite regular adherence to inhalatory hypertonic saline therapy and respiratory physiotherapy. *Pseudomonas aeruginosa* (Pa) has never been isolated from her sputum and her serum anti-*Pseudomonas aeruginosa* IgG antibodies are low.

3.2. Genetic analyses

Sequencing in the patient revealed the c.3140-26A>G *CFTR* variant (reference sequence NM_000492.3) in homozygous state; no additional disease causing variants were identified in the coding region or in the exon/intron boundaries. Parents were shown to be carriers of the same variant in heterozygous state. *CFTR* mRNA analysis showed an aberrant splicing pattern with 25 nucleotides inserted between exons 17A in 17B in the patient but not in her mother nor in healthy control subjects. The 25-nucleotide insertion was matching the last 25 nucleotides of intron 17a (namely TGTTTTCTATGGAAATATTTTCACAG) as previously reported.¹⁵

3.3. Comparison of clinical data of patients with the c.3140-26A>G *CFTR* variant listed in the ECFSPR and the CFTR2 database and our patient

After the c.3140-26A>G variant was confirmed in our patient, clinical data on patients with this variant was requested from the ECFSPR according to standard regulations. There are 195 patients with the c.3140-26A>G mutation currently listed in the ECFSPR. 191 (97.95%) are heterozygous and 4 (2.05%) homozygous for this variant. At the last data review, all 4 ECFSPR patients were alive. 2 were younger and 2 older than 18 years of age. Their mean age was 20 years, the youngest 3.3 and oldest 34.3 years old. Their mean age at diagnosis was 6 years, the earliest at 3 months and the latest at almost 13 years of age. 2 were diagnosed before the first year of life, none before the first month.

Their lung function described as mean percent of predicted FEV1 was 77.23% for the patient younger than 18 years and 89.77% for the 2 patients older than 18 years. The lung function of the 3 year old could not be measured yet by standard spirometry.

Patients over 18 years of age had a mean BMI of 26.22 kg/m² and the BMI expressed with the mean Z score for the two patients under 18 was -0.18, with a minimum of -1.13 and maximum of 0.76.

Only one patient needed pancreatic enzyme replacement therapy, the other 3 were pancreatic sufficient. 1 patient was chronically colonized with *S. aureus* and probably none with *P. aeruginosa*, but the information on Pa is missing for one patient. The clinical data are summarized in Table 2.

Table 2. Comparison of clinical data of patients with the c.3140-26A>G *CFTR* variant in homozygous state listed in the ECFSPR with the data of our patient.

Clinical variable	ECFSPR patients (Total number: 4)			Our patient
	mean	minimum	maximum	
Age (years)	20	3.3	34.4	24
Age at diagnosis (years)	6	0.25	12.91	16
FEV1 (percent predicted) <18 years	77.23	77.23	77.23	/
FEV1 (percent predicted) >18 years	89.77	85.25	94.3	88
BMI (Z score) < 18 years	-0.18	-1.13	0.76	
BMI (kg/m ²) > 18 years	26.22	22.77	29.67	20

	no	yes	missing data	
Use of pancreatic enzymes	3	1	0	no
Chronic <i>Pseudomonas aeruginosa</i>	3	0	1	no
Chronic <i>Staphylococcus aureus</i>	3	1	3	yes

Clinical features of homozygous patients with the c.3140-26A>G variant were compared to all the other CF patients with 2 CF causing variants listed in the CFTR2 mutation database and to the here reported patient. The data are presented in Table 3.

Table 3. Clinical features of the c.3140-26A>G homozygous patients as listed in the CFTR2 mutation database, compared to all listed CF patients with 2 disease causing mutations and to our patient.

Clinical Feature	Average of all patients homozygous for c.3140-26A>G	Our patient	Average of all patients with 2 disease causing mutations in the database
Sweat chloride value (mmol/l)	90	57	96
Lung function (FEV1% predicted)	<10 years: insufficient data 10-20 years: insufficient data >20 years: insufficient data	70%	<10 years: 63-124% 10-20 years: 42-118% >20 years: 25-104%
Pancreatic status	33% (2 patients) insufficient	sufficient	85% (66.394 patients) insufficient
<i>Pseudomonas aeruginosa</i> chronic colonization	33% (2 patients)	No	55% (34.460 patients) chronically colonized
Average age in years	16	24	20

4. Discussion with review of the published literature

A review of the literature in search for the variants characteristics in terms of disease severity was done. A PubMed search using keywords 3272-26A>G or c.3140-26A>G and cystic fibrosis yielded 6 articles. References of relevant articles were analyzed and further 7 articles that mention the mutation were identified. The found references in chronological order are shown in Table 4.

Table 4. Clinical characteristics of the in the published literature found reports of patients with the c.3140-26A>G mutation and chronological characterization of the variant.^{14-16, 20-23}

First author, year of publication	Number of patients with c.3140-26A>G	Genotype: c.3140-26A>G in combination with	Phenotype						Variant characterization
			Mean age	Mean age at diagnosis	Mean sweat chloride test	Pancreatic function	Nasal polyps	Mean FEV1	
Fanen et al, 1992 ²³	1	p.W846X	No data	No data	No data	Sufficient	No data	Mild pulmonary disease	
Kanavakis et al, 1995 ¹⁶	3	2: p.Phe508del 1: unknown	11 years (SD 7)	9.5 years (SD 5)	105mEq/l (SD 7.6)	All pancreatic sufficient	2/3	108% (SD 22.6)	Proposition of the splicing defect.
Beck et al 1999 ¹⁵	5	5: p. Phe508del	18 years (SD 10.4)	9 years (SD 13.9)	103mEq/l (SD 7.3)	4/5 sufficient, 1/4 mild insufficiency	4/5	68% (SD 24.5)	mRNA confirmation of the splicing defect. Mutated protein detected on the epithelial cell membrane.
Amaral et al, 2001 ¹⁴	60	44: p. Phe508del 16: other	20,5 years (SD	8 years (SD 20)	99,5mEq/l (SD 47)	39 sufficient, 16	19/60	87% (SD 47)	Additional report of the mutation.

		disease causing	17.5)			insufficient			
Feldmann et al, 2003 ²⁰	2	2: p. Phe508del	27 years (SD 10)	No data	62.5mEq/l (SD 2)	No data	No data	No data	Additional report of the mutation.
Storm et al, 2007 ²¹	11	10: p. Phe508del 1: c.1717-1G>A	Clinical phenotypes milder than p. Phe508del homozygotes.						Additional report of the mutation.
Jung et al, 2011 ²²	1	1: p.Phe508del	No data						Additional report of the mutation.

211

212 To our knowledge, this is the first longitudinal clinical report on a patient with this variant in
213 homozygous state. According to the so far published literature, c.3140-26A>G is a rare
214 variant so far detected mostly in compound heterozygosity with other disease causing
215 variants. Patients usually have mild disease even when this mutation is paired with a severe
216 disease causing variant. The patients are mostly pancreatic sufficient, have good lung function
217 are on average older and diagnosed at an older age than patients with classic CF variants, but
218 have elevated sweat chloride levels and a higher incidence of nasal polyps.^{15-17, 20-23}

219 The largest number of patients with c.3140-26A>G in heterozygous state was presented in a
220 European study by Amaral et al in 2001.¹⁴ The study included 60 patients from 6 European
221 countries: France, Spain, Greece, Germany, Portugal and Belgium. The mean age of the
222 patients at the time of the study was 20.5 (SD 17.5) years. 44 of the patients had the c.3140-
223 6A>G and the p. Phe508del (c.1521_1523delCTT) combination of disease causing alleles and
224 16 had c.3140-26A>G in combination with another CF causing variant. Their mean age at
225 diagnosis was 8 (SD 20) years, sweat test chlorides 99.5 (SD 24.7) mEq/l and FEV1 87 (SD
226 47) % predicted. 19 had nasal polyposis and 18 Pa colonization.¹⁴ The other authors reported
227 patients from individual countries or centers and therefore had lower numbers of described
228 patients, all patients were heterozygotes for the c.3140-26A>G variant.

229

There are two ways to assess the severity of a mutation – on the basis of a patient’s clinical status and on a molecular basis. The first is the most reliable but requires long term follow-up. The second can be used as a prediction tool for an approximate evaluation of the probable clinical findings to come. Based on the found data in the literature and the clinical status of our patient we were interested if there is a difference in mutated mRNA translation between hetero and homozygotes for the c.3140-26A>G mutation compared to healthy controls. On the molecular level, the effect of a gene variant can be studied by observing its influence on RNA translation and protein synthesis. Splicing is a process in which a premature mRNA transcript is formed into mature mRNA. In this process introns are removed and exons which are the coding nucleotide sections within a gene are joined together into mature mRNA which is then translated into a protein.²⁴ Errors can occur in splicing, as in all processes. There are presumed to be three classes of variants that effect splicing: variants of a splice site that cause loss of function of that site, variants that cause a disruption of the reading frame and displacement of a splice site that causes shorter or longer exons to be formed.²⁴ The HGMD professional 2019.3 currently reports more than 240 different intronic disease causing variants in the *CFTR* gene. 6 of them are located in the 3' splice site of intron 17A, among them 4 including the here reported c.3140-26A>G in a polypyrimidine tract located between the branch site and the basic acceptor splice site.⁷ The c.3140-26A>G *CFTR* variant was first described by Fanen et al in 1992 and was suspected to be a variant that effects mRNA splicing.²³ Additional reports followed in 1995 by Kanavakis et al and in 1999 by Beck et al where aberrant splicing was demonstrated on the mRNA level.^{16, 15} The c.3140-26A>G variant results in an insertion of 25 nucleotides of intron 17a of the *CFTR* gene as confirmed in our patient and causes a frameshift and a premature stop codon in exon 17b.¹⁵ The same insertion in mRNA was present in here reported patient with, They proposed that some of the alternative transcript reaches the cell membrane as a modified CFTR protein. Because of the location of the insertion the alternative protein retains the wild-type CFTR sequence until amino acid residue 1046 which could enable it to have some single chloride conductance remaining. It was proposed that the 36 amino acid extension caused by the insertion can have three different effects on CFTR folding, trafficking and function: a change in stability, missed interaction with PDZ-domain proteins with a probable influence on CFTR stability, chloride conductance regulation and its localization or that the protein would not fold correctly.¹⁶ In 2001 Amaral et al reported that the alternative acceptor splicing site in intron 17a competes with normal *CFTR* mRNA.¹⁴ Therefore, the remaining normal mRNA is normally processed and leads to some remaining normal protein at the cell membrane. It was postulated that the

remaining normal *CFTR* mRNA still existing in patients with the c.3140-26A>G variant even when paired with a severe disease causing mutation lessens the severity of CF disease. It was hypothesized that the c.3140-26A>G variation on both alleles would avoid CF totally.¹⁵ This was not confirmed in our study, since the homozygous patient had only mutated splicing pattern present in nasal epithelial cells and normal transcript was not detected. Furthermore, in the analysed heterozygous mother, only normal splicing was present, probably due to preferential transcription of the normal allele. Of course, *CFTR* expression is tissue specific and the splicing pattern was analysed in nasal epithelial cells, where *CFTR* gene is known to be expressed in lower levels.^{25, 26} A mild disease presentation in patients carrying nonsense mutations was previously explained by the presence of alternative spliced mutant transcripts.²⁶ This could also be the case in the c.3140-26A>G variant, since we have studied only a selected part of the mRNA transcript in nasal epithelial cells. The other possibility is a presence of spliceosome-mediated RNA trans-splicing already proven to functionally correct endogenous *CFTR* mutant protein.²⁸

5. Conclusions

Incredible progress has been made in the past 5 years in the treatment of CF. New gene variant specific medication with *CFTR* modulators has improved the life and outcome of patients with specific gene defects. Currently numerous *CFTR* variants are known, but the clinical significance of most of them remains trivial. Case reports on patients with rare *CFTR* variants are therefore beneficial for the prediction of the clinical course of the disease in other patients with the same variant. 12 year longitudinal data on a CF patient with the c.3140-26A>G variant in homozygous state that is presented in this report adds new knowledge. In our study, aberrant splicing of the mRNA due to the c.3140-26A>G variant was shown, probably resulting in the mild clinical presentation of the disease. Only speculations can be made on the future quality of life of our patient, but based on the current data we might predict a good outcome especially with the implementation of new variant specific therapy into everyday patient care.

6. Acknowledgements

We would like to thank the people with CF and their families for consenting to their data being included in the European Cystic Fibrosis Society Patient Registry, the ECFSPR for

providing access to patient data and the individual country representatives for allowing the use of data for publication (www.ecfs.eu/projects/ecfs-patient-registry/steering-committee).

7. References

1. P. M. Farrell, B. J. Rosenstein, T. B. White, F. J. Accurso, C. Castellani, G. R. Cutting, et al, *The Journal of Pediatrics*. **2008**, 2: S4–S14. DOI: 10.1016/j.jpeds.2008.05.005
2. A. Kotnik Pirs, M. Praprotnik, K. Trebusak Podkrajsek, K. Seme, M. Aldeco, D. Lepej, J. Breclj, N. Bratanic, U. Krivec, *Slov. Pediatr*. **2014**, 21: 164–171.
3. B. Kerem, J. M. Rommens, J. A. Buchanan, D. Markiewicz, T. K. Cox, A. Chakravarti, M. Buchwald, L. C. Tsui, *Science*. **1989**, 245: 1073–1080.
4. K. De Boeck, M.D. Amaral. *Lancet Respir. Med*. **2016**, 4(8): 662-674, DOI: 10.1016/S2213-2600(16)00023-0
5. J. Zielinski, L. C. Tsui. *Annu. Rev. Genet*. **1995**, 29: 777-807.
6. K. De Boeck, M. Wilschanski, C. Castellani, C. Taylor, H. Cuppens, J. Dodge, M. Sinaasappel, Diagnostic Working Group, *Thorax*. **2006**, 61: 627–635.
7. HGMD professional 2019.3, <http://www.hgmd.cf.ac.uk/ac/index.php>
8. D. H. Andersen. *Am. J. Dis. Childhood*. **1938**, 56, 344-399.
9. R. S. Pettit, C. Fellner. *P T*. **2014**, 39(7), 500-511.
10. K. De Boeck, N. Derichs, I. Fajae, H. R. de Jonge, I. Bronsveld, I. Sermet, F. Vermeulen, D. N. Sheppard, H. Cuppens, M. Hug, P. Melotti, P. G. Middleton, M. Wilschanski on behalf of the ECFS Diagnostic Working Group and EuroCareCF WP3 Group on CF diagnosis. *J Cyst. Fibros*. **2011**, 10(Suppl 2), S53–S66.
11. C. Castellani, H. Cuppens, M. Macek Jr, J. J. Cassiman, E. Kerem, P. Durie, et al. *J Cyst. Fibros*. **2008**, 7, 179–196.
12. Cystic Fibrosis Mutation Database. Available at [http:// www.genet.sickkids.on.ca](http://www.genet.sickkids.on.ca)
13. The Clinical and Functional Translation of CFTR (CFTR2); Available at <http://cftr2.org>
14. M. D. Amaral, P. Pacheco, S. Beck, et al. *J Med. Genet*. **2001**, 38, 777–782.
15. S. Beck, D. Penque, D. Garcia, A. Gomes, C. Farinha, L. Mata, S. Gulbenkian, K. Gil-Ferreira, A. Duarte, P. Pacheco, C. Barreto, B. Lopes, J. Cavaco, J. Lavinha, M. D. Amaral. *Human mutation* **1999**, 14, 133–144.
16. E. Kanavakis, M. Tzetis, T. Antoniadis, J. Trager-Synodinos, C. Kattamis , S. Doudounakis, G. Adam. *J Med. Genet*. **1995**, 32, 406–407.

- 329 17. P. R. Sosnay, K. R. Siklosi, F. Van Goor, K. Kaniecki, H. Yu, N. Sharma, A. S.
330 Ramalho, M. D. Amaral, R. Dorfman, J. Zielenski, D. L. Masica, R. Karchin, L. Millen, P. J.
331 Thomas, G. P. Patrinos, M. Corey, M. H. Lewis, J. M. Rommens, C. Castellani, C. M.
332 Penland, G. R. Cutting. *Nat. Genet.* **2013**, 45, 1160–1167.
- 333 18. E. Dequeker, M. Stuhmann, M. A. Morris, T. Casals, C. Castellani, M. Claustres, H.
334 Cuppens, M. des Georges, C. Ferec, M. Macek, P. F. Pignatti, H. Scheffer, M. Schwartz, M.
335 Witt, M. Schwarz, E. Girodon. *Eur J Hum. Genet.* **2009**, 17, 51–65.
- 336 19. A. R. Smyth, S. C. Bell, S. Bojcin, M. Bryon, A. Duff, P. Flume, N. Kashirskaya, A.
337 Munck, F. Ratjen, S. J. Schwarzenberg, I. Sermet-Gaudelus, K. W. Southern, G. Taccetti, G.
338 Ullrich, S. Wolfe. *J Cyst. Fibros.* **2014**, 13 (Suppl 1), S23–S42.
- 339 20. D. Feldman, R. Couderc, M. P. Audrezet, C. Ferec, T. Bienvenu, M. Desgeorges, M.
340 Clusters, H. Mitre, M. Blayau, D. Bozon, et al. *Hum. Mutat.* **2003**, 22, 340.
- 341 21. K. Storm, E. Moens, L. Vits, H. De Vlieger, G. Delaere, M. D’Hollander, W. Wuyts, M.
342 Biervliet, L. Van Schil, K. Desager, M. M. Nothen. *J Cyst. Fibros.* **2007**, 6, 371–375.
- 343 22. H. Jung, C. S. Ki, W. J. Koh, K. M. Aho, S. I. Lee, J. H. Kim, J. S. Ko, J. K. Seo, S. L.
344 Cha, E. S. Lee, J. W. Kim. *KJLM.* **2011**, 31, 21924.
- 345 23. P. Fanen, N. Ghanem, M. Vidaud, C. Besmond, J. Martin, B. Costes, F. Plassa, M.
346 Goossens. *Genomics.* **1992**, 13, 770–776.
- 347 24. K. H. Lim, L. Ferraris, M. E. Filloux, B. J. Raphael, W. G. Fairbrother. *Proc. Natl.*
348 *Acad. Sci. USA.* **2011**, 108, 11093–11098.
- 349 25. B. C. Trapnell, C. S. Chu, P. K. Paakko, et al. *Proc. Natl. Acad. Sci. USA.* **1991**, 88,
350 6565–6569.
- 351 26. N. L. White, C. F. Higgins, A. E. Trezise. *Hum. Mol. Genet.* **1998**, 7, 363–369.
- 352 27. A. Hinzpeter, A. Aissat, E. Sondo, C. Costa, N. Arous, C. Gameiro, N. Martin, A.
353 Tarze, L. Weiss, A de Becdelievre, et al. *PLoS Genet.* **2010**; 6. pii: e1001153. DOI:
354 10.1371/journal.pgen.1001153
- 355 28. X. Liu, Q. Jiang, S. G. Mansfield, M. Puttaraju, Y. Zhang, W. Zhou, J. A. Cohn, M. A.
356 Garcia-Blanco, L. G. Mitchell, J. F. Engelhardt. *Nat. Biotechnol.* **2002**, 20, 47–52.
- 357