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Scientific paper

Sandpaper Wastes as Adsorbent for the Removal of Brilliant Green and Malachite Green Dye

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Abstract

Sandpaper wastes were used as adsorbent after pyrolysis at 500 °C and calcination at 800 °C for the removal of brilliant green and malachite green cationic dye from an aqueous solution. The effects of the pH, the adsorbent dose, the contact time, and the initial dye concentration on the removal efficiencies were investigated. The isotherm studies were conducted by using the Langmuir, Freundlich, and Dubinin-Radushkevich models, and thermodynamic studies were also performed. The adsorption of the Brilliant green and malachite green were found to comply with the Langmuir isotherm model and the Freundlich isotherm model, respectively. The thermodynamic studies showed that the adsorption of dyes were endothermic. The E values obtained from the Dubinin-Radushkevich isotherm showed that the adsorption mechanism was chemical in nature. Furthermore, the three kinetic models (pseudo first-order, pseudo second-order, and intraparticle diffusion) were investigated. It was found that the pseudo second-order kinetic model fitted well for adsorption of dyes.

Keywords: Adsorption; brilliant green; malachite green; dye removal; sandpaper.

1. Introduction

Brilliant green and malachite green are cationic (basic) dyes. Many industries such as paper, textile, furniture, and food industries use dyes for coloring purposes.[1,2] Furthermore, in fish farming, brilliant green is used to protect fish from fungi, from parasites and from infections. However, the consumption of fish produced in this way is not recommended.[2–5] Cationic dyes have toxic, carcinogenic and mutagenic properties.[6] Significant risks arise after exposing these dyes to people. Therefore, the removal of the dyes prior to their discharge into the environment is crucial and essential.

Physical, chemical and biological methods are widely used for the removal of dyes from water. Among these methods, adsorption, the physical method has advantages such as simplicity, low cost, and ease of application. Natural materials (raw or activated forms of clay minerals), synthesized materials, nanomaterial based adsorbents, agricultural wastes and by-products (raw or modified leaf based materials, coffee wastes, peels) and industrial wastes and by-products (fly ash, aluminum oxides), and activated carbon are the most used adsorbents for dye removal.[1,7–11] Cost is an important parameter for choosing the adsorbent. Low-cost adsorbents include natural, agricultural and industrial by product wastes[12]. Furthermore, the waste materials have little or no economic value and usually present a disposal problem.[7] The use of these waste materials for the purpose of wastewater treatment can play a significant role in solving the disposal problems. Numerous inexpensive and abundant biosorbents especially agro waste materials, as well as industrial and municipal wastes, have been proposed by several researchers for the removal of malachite green and brilliant green dyes from aqueous solution.[13] The usage of waste as an adsorbent helps to reduce environmental pollution by recycling. In the literature some of the low-cost adsorbents used for dye removal were NaOH treated saw dust,[14] waste rubber tire,[15] white rice husk ash,[16] Neem leaf powder,[17] kaolin,[2] peach stone,[18] and medical cotton waste[19] etc.

Sandpaper is an abrasive used in the sanding process to correct the rough surfaces. It consists of sheets of paper or cloth with abrasive material glued with resin to one face. Formerly, sand and glass were used as abrasive surfaces, but nowadays materials such as aluminum oxide, zirconium oxide, and silicon carbide etc. are used.[20] The storage and disposal of the sandpaper waste is a problem in terms of time, space, and cost. With this work, a useful area for sandpaper waste has been created, which will be beneficial for the environment and waste water remediation.

The aim of the study is the removal of brilliant green and malachite green dyes from water by using sandpaper waste. Two adsorbents were prepared by applying pyrolysis and calcination process. To the best of our knowledge, it is the first study on brilliant green and malachite green removal using sandpaper waste based adsorbent. The important point of the study is that the pollution is reduced both by recycling of sandpaper waste, which is an industrial waste material and by removing the dyes from water. The effects of the experimental parameters such as the pH, the adsorbent dose, the contact time and the initial dye concentration were examined. The isothermal models (Langmuir, Freundlich and Dubinin-Radushkevich), thermodynamic and kinetic parameters (pseudo first order, pseudo second order and intraparticle model) were also evaluated.

2. Experimental

2. 1. Materials and Apparatus

All the reactive used was of an analytical grade. Distilled water was used throughout the study. Cationic dye Brilliant Green (BG) (CI 42040, MW: 462.65), malachite green oxalate (MG) (CI 42000, MW: 927.01) hydrochloric acid, sodium hydroxide, acetic acid, and sodium acetate were obtained from Merck. The chemical structure of brilliant green and malachite green are presented in Figure 1. The working dye solution was prepared daily by diluting 1000 mg/L stock dye solution. The spectrophotometric measurements were carried out by TG 80+ model double beam UV/Vis spectrophotometer with PG Instruments. The pH was measured using a Mettler Toledo Five Go FG-2 pH meter. A Biosan OS-10 orbital shaker at 350 rpm and Nuve ST-402 vibration water bath were used for the adsorption studies. The FTIR analyses of the adsorbents were carried out by using the Perkin Elmer 100 spectrum FT-IR spectrometer in the range of 4000–400 cm^{-1}. The pore and surface morphology images were captured by using the Thermo Scientific Apreo S LoVac model scanning electron microscope (SEM). The chemical compositions of SW500 and SW800 were analyzed by X-ray Fluorescence spectrometer (Spectro Xepos, Ametec). BET analysis were executed by Quantachrome ASiQwin. The sandpaper was supplied by a fibre disk pad production company in Izmir, Turkey, as sheets.

2. 2. Preparation of Adsorbent from Sandpaper Wastes

Two kind of adsorbent were obtained after pyrolysis and calcination processes. Pyrolysis and calcination processes were separately applied to the sandpaper sheets. The sandpaper sheets were cut into small pieces (≤ 2 cm) before the experiments. For pyrolysis, firstly, a quantity of 50 g of sandpaper was loaded into the reactor, and then the reactor was heated with a temperature rate of 7 °C per minute up to 500 °C and held at this temperature for 1 h. The reactor was continually purged with nitrogen at a flow rate of 25 mL/min. The nitrogen gas swept the volatile products from the reactor into the ice-cooled traps. The condensable volatiles, which were collected in the traps, were released. After pyrolysis, the furnace was cooled to room temperature in a nitrogen gas stream and the reactor content (carbonized residue) was withdrawn from the reactor.[18] The obtained adsorbent was named SW500. Then, another part of the sandpaper sheets was calcined at 800 °C in a furnace up to 16 h and then stored in desiccators. The obtained adsorbent was named SW800.

2. 3. Adsorption Studies

All of the adsorption studies were examined in batch mode. In order to find out the optimum experimental conditions, 25 mL of dye solutions were used. The initial concentration used was 20 mg/L. The contact time was 24 h unless otherwise stated. In the pH study, the adsorbent amounts were 10 mg for both adsorbents. The optimal sorbent doses were found to be 1.2 g/L of SW500 and 0.4 g/L of SW800 for the BG removal, while 2.4 g/L for SW500 and 0.6 g/L for SW800 for the MG removal after optimization study. The pH effect for the removal efficiencies was studied in the pH range between 3 and 10. The initial pH of the solutions was adjusted to the desired value using NaOH or HCl. The optimization studies, such as adsorbent dose (0.2, 0.4, 0.6, 1.2, 1.8 and 2.4 g/L), contact time (1, 5, 10, 15, 20, 40, 60, 120, 180, 240, and 1440 min), ini-

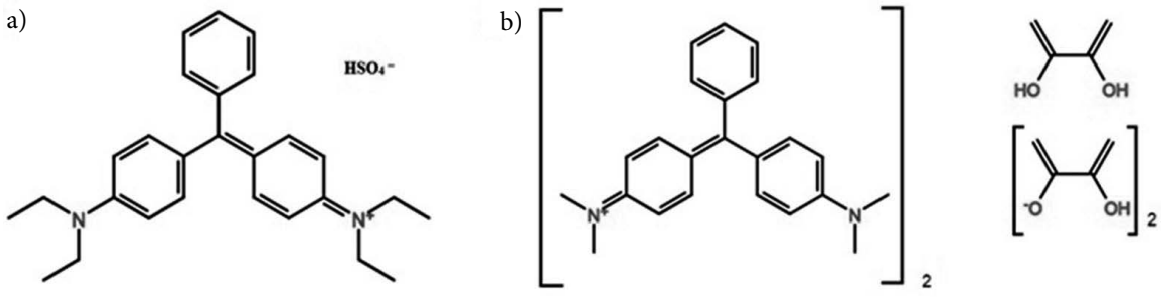


Figure 1. The chemical structure of brilliant green (a) and malachite green (b)

tial dye concentrations (5, 10, 25, 50, 100, 200, and 400 mg/L), and temperature (298 K, 303 K, 313 K, and 323 K) were performed. The adsorption isotherms were evaluated in the range of 5–500 mg/L of dyes. Kinetic studies were investigated between 1–1440 min. The remaining BG and MG dye concentrations after sorption were measured at 624 nm and 617 nm by using UV-Vis spectrophotometer, respectively. All the experiments were conducted in triplicate. Before the spectrophotometric measurements, the pH of the dye solutions and standard solutions for the calibration were adjusted to 5.5 by using an acetic acid/acetate buffer. The removal efficiencies (R, %) and adsorbed dye amounts (q, mg/g) were calculated, respectively;

$$R(\%) = \frac{(Ci-Ce)}{Ci} \times 100 \quad (1)$$

$$q = \frac{(Ci-Ce)}{W} \times V \quad (2)$$

Here, Ci and Ce are dye concentrations at an initial and equilibrium (mg/L), w is the amount of the adsorbent (g), and V is the volume of the dye solutions (L).

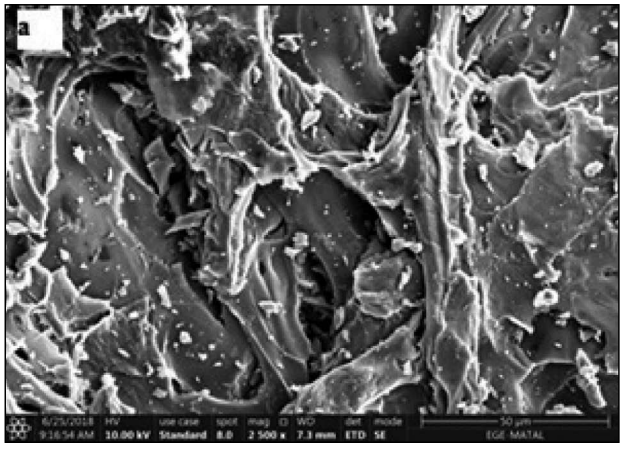
3. Results and Discussion

3. 1. Characterization of the Sandpaper Waste Adsorbent

The morphology of the bare SW500 and SW800 are depicted in Figure 2a and 2b, respectively. The morphology of the SW500 surface was irregular and porous. In Figure 2b, it was seen that the particles of SW800 were spherical and aggregate. The higher number of pores increased the adsorption of the dyes.

The FTIR spectra of the adsorbents before and after dye adsorption are shown in Figure 3a and 3b. In the spectrum of SW800, the peak at 3454.78 cm^{-1} could indicate –OH stretching of the phenolic structure and crystal water. The intense peak at 1439.07 cm^{-1} was attributed to the aliphatic C–H stretching band. The peaks belonging to the crystal water and aliphatic C–H stretching band could also be seen in the other spectra. The two bands at 873.67 and 577.56 cm^{-1} could be Al–O vibration bands in Al_2O_3. After the adsorption for both the adsorbents, the C = C bands belonging to the aromatic ring of BG and MG appeared in the range of 1600–1700 cm^{-1}. After BG adsorption on SW500, a peak assigning N–C band at 2969 cm^{-1} appeared. Besides, several adsorption peaks that emerged in the range of 1550–1380 cm^{-1} might be ascribed to the N–C groups after BG adsorption on SW500.[13,14,21–26]

The chemical compositions of SW500 and SW800 were determined by X-ray fluorimeter. It was found that SW500 contained 1.43% Al_2O_3, 0.31% SiO_2, 0.72% P_2O_5, 0.16% SO_3, 12.47% CaO, 0.68% TiO_2, 0.74% Fe_2O_3, 0.03% CuO, 0.24% ZnO, 0.03% SrO and 0.05% ZrO_2, while SW800 contained 3.6% Al_2O_3, 0.97% SiO_2, 0.67% P_2O_5, 0.16% SO_3, 44.06% CaO, 1.27% TiO_2, 1.99% Fe_2O_3, 0.01% CuO, 0.03% ZnO, 0.08% SrO and 0.09% ZrO_2. The amount of CaO in SW800, which was prepared by calcination, was found higher than that of SW500. Furthermore, higher dye removal efficiencies were obtained for SW800 in the adsorption studies. Therefore, that result could be a conclusion of existence of higher amount of CaO in SW800. Metal oxides containing calcium oxides are well known adsorbents for removal of various effluent gas streams. Because they have high adsorption capacity, high surface reactivity, low cost, and abundant.[27] Calcium mineral is also efficiently used for the dye removal in the literature. Calcium rich biochar from crab shell showed highly efficient removal for Malachite Green and Congo Red although it showed low specific surface area and total pore volume.[28] Jung et al. have synthesized an adsorbent using spent coffee grounds (SCG) calcium alginate beads for the removal of acid orange 7 and methylene blue. It was expressed that it was difficult to remove powdered SCG-based activated



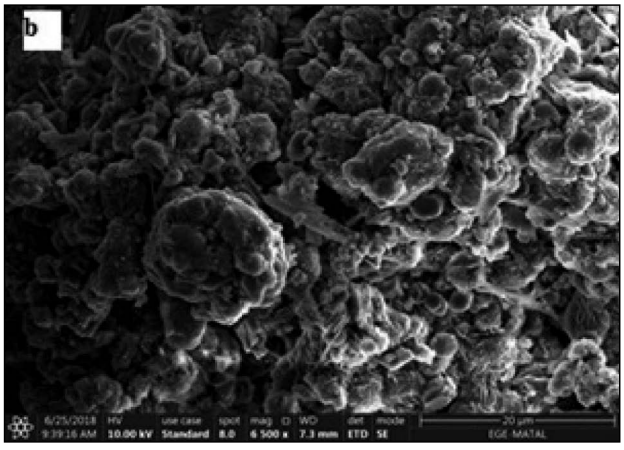


Figure 2. SEM images of bare adsorbents a) SW500 and b) SW800

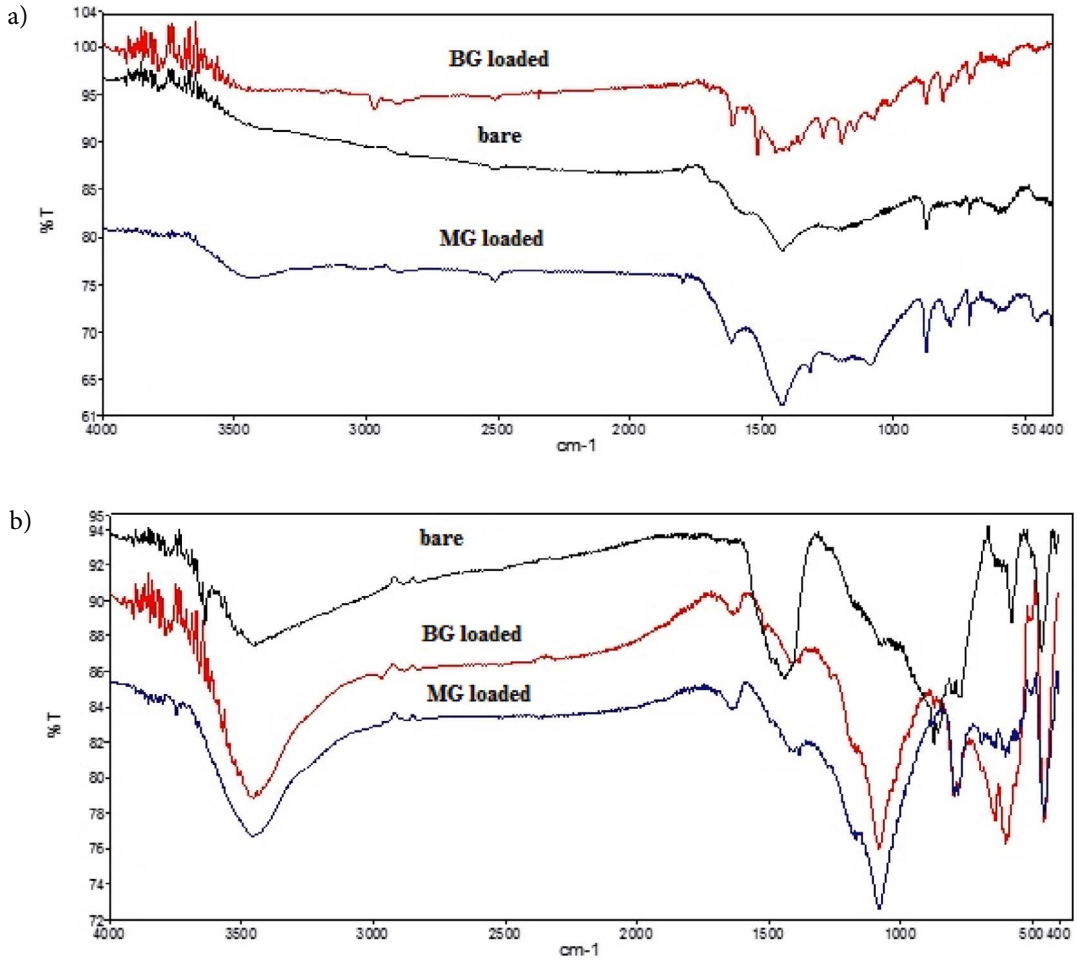


Figure 3. FTIR spectra of adsorbents before and after dye adsorption a) SW500 and b) SW800

carbon from aqueous solution after adsorption. Therefore, to form porous hydrogels beads powdered SCG based activated carbon was entrapped in calcium-alginate beads. It was reported that such heterogeneous surface might be take part in removal of dyes.[29] Basic Green 4 was successfully removed by using sea shell powder. It was reported the shell contained protein, calcite and calcium carbonate crystals, and the adsorbent had heterogeneous pores and cavities that gave large surface area for dye removal.[30] In study of Xia et al., it was expressed that the various metal oxides for the adsorption capacities of Congo red dye were increased in the following order: NiO< MnO_2<Cr_2O_3< Fe_2O_3<MgO<CaO.[27] Aguayo-Villarreal et al. clearly mentioned that the adsorption of acid blue 74, acid blue 25 and reactive blue 4 was governed by the calcium compounds existing in pecan shells. The electrostatic interactions between calcium ion and the sulphonyl groups of the dyes molecules were thought to be responsible for adsorption of dyes on pecan shells.[31]

The BET surface area, total pore volume and pore size were determined as 3.070 m^2/g, 0.004354 mL/g, 28.37 A° for SW500, and 1.103 m^2/g, 0.001933 mL/g, 35.04 A° for SW800, respectively.

3. 2. Adsorption Studies for BG and MG Removal

3. 2. 1. Effect of pH on BG and MG Removal

In order to define the optimum adsorption pH, the pHs of the solutions were set in the range of 3–10 by 0.01 mol/L HCl and NaOH. The removal efficiencies are shown in Figure 4. The optimum pH range of BG was found to be between 3 and 10, and the optimum pH of MG was in the range of 4–10 for SW800. The removal efficiencies of BG were reached to 96% at pH 5, while the removal efficiencies of MG reached 91.4% at pH 7 for SW 500. The point of zero charge (pH_{pzc}) was determined according to the following procedure.[32] 25 mL of 0.1 mol/L of KNO_3 solution was adjusted to different pH values using HCl or NaOH and was added to the adsorbents. Thereafter, the suspension was shaken for 24 h to obtain the equilibrium pH. The change of the pH during the equilibrium was calculated by subtracting the initial pH values from the final pH values. The ΔpH values were then plotted against the initial pH values. The initial pH at which the ΔpH was zero was taken to be the pH_{pzc}. The pH_{pzc} values were 7.30 for SW500 and 10.1 for SW800. At

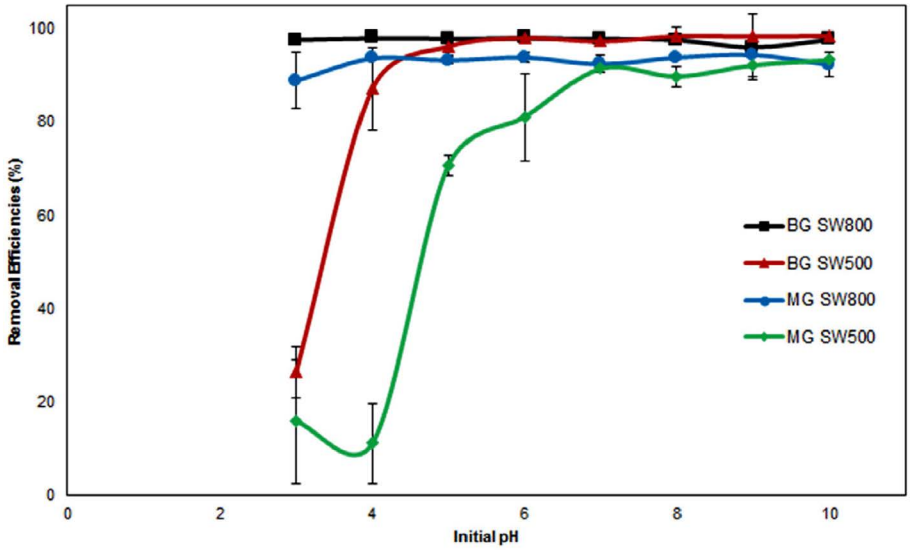


Figure 4. Effect of initial pH on the removal of BG and MG (adsorbent amount 10 mg, initial dye concentration 20 mg/L, volume 25 mL, contact time 24 h, pH range 3–10, n = 3)

the pH values higher than pHpzc, the surface charge was negative and attracted positively charged dye while at lower pH values, the surface charge was positive and attracted negatively charged dye.[32,33] It was found that the final pH of the solutions at the end of the adsorption was found to be 7.5 after the initial pH of 5 for SW500, and about 10.8 for SW800 in all the studied initial pHs., Before the pH_{pzc}, the adsorption efficiencies of BG and MG for SW500 were as low as expected because of electrostatic repulsion. As seen in Figure 4, the adsorption efficiencies of BG were above 98% for SW500 after the initial pH of 5 (final solution pH 7.5 for SW500). In addition, the low adsorption observed for SW500 at pH below 5 may be due to the competition between H^+ ions and dye cations for the adsorbent's active sites.[34] Furthermore, the same trend was observed for MG removal. Therefore, it was thought that electrostatic forces were effective for BG and MG removal. SW800 provided a wider working range than that of SW500 for BG and MG removal.

3. 2. 2. Effect of Adsorbent Dose on BG and MG Removal

The effect of the sorbent dose was investigated in the range of 0.2–2.4 g/L. The results are presented in Figure 5. As seen in the Figure, for SW800, the removal efficiencies of BG in the studied range did not change significantly; therefore, the optimum dose was selected as 0.4 g/L. For SW500, the removal efficiencies of BG and MG increased slightly and then reaching a constant value of 1.2 g/L and 2.4 g/L, respectively. The optimum sorbent dose was found to be 0.6 g/L for MG removal for SW800. That situation

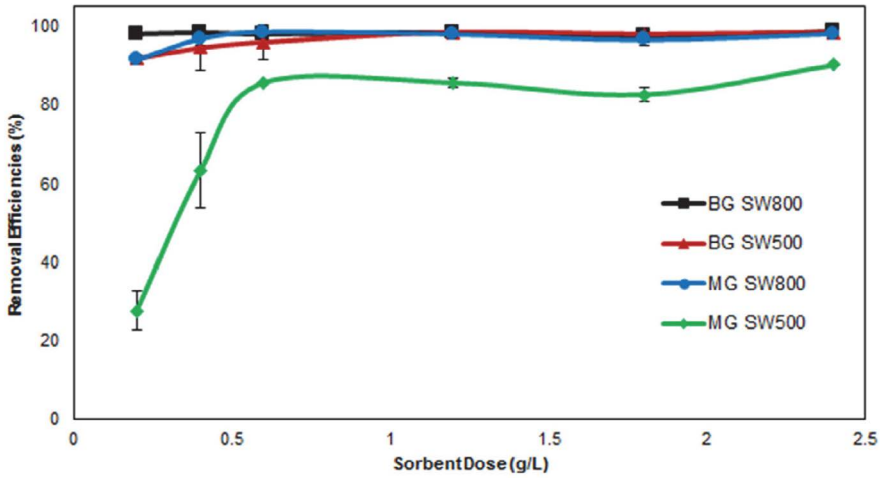


Figure 5. Effect of adsorbent dose on the removal of BG and MG (adsorbent dose range 0.2–2.4 g/L, initial dye concentration 20 mg/L, volume 25 ml, contact time 24 h, n = 3)

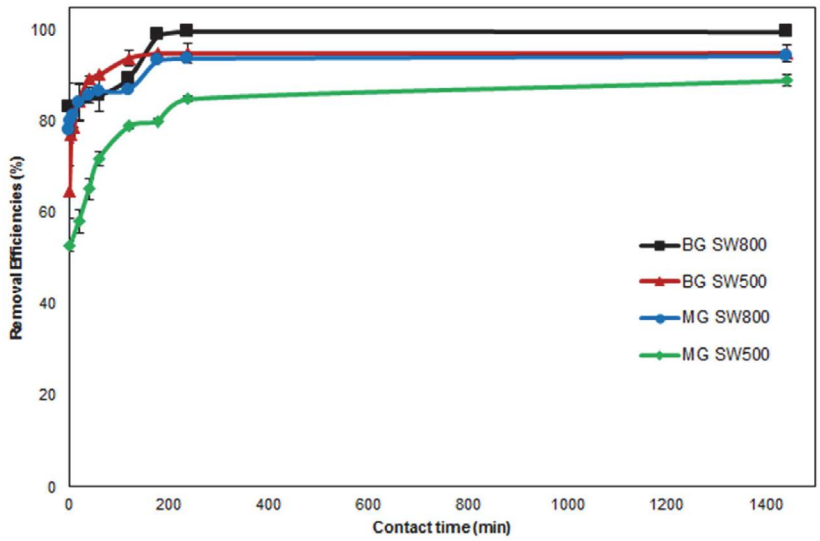


Figure 6. Effect of contact time on the removal of BG and MG (adsorbent dose of BG and MG: 0.4 and 0.6 g/L for SW800 and, 1.2 and 2.4 g/L for SW500, initial dye concentration 20 mg/L, volume 25 mL, contact time 1–1440 min, n = 3)

may be attributed to an increase in the number of active sites with the increase in the adsorbent dose[2].

3. 2. 3. Effect of Contact Time on BG and MG Removal

Figure 6 shows the effect of the contact time on the removal of BG and MG by SW500 and SW800 adsorbents in the range of 1–1440 min. It was observed that the adsorption equilibrium is reached faster for SW500 than for SW800 for BG removal. Within the first 40 minutes, 89% of BG dye was adsorbed by SW500 and reached 94% of the removal efficiency at 120 min. Meanwhile, 90% of BG dye adsorption took place within 120 min and equilibrium was reached at 180 min with 98.8% of the removal efficiency for SW800. The optimum contact times of BG dye were selected to be 120 min and 180 min for SW500 and SW800, respectively. The optimum contact times of MG removal were found to be 180 min and 240 min for SW500 and SW800, respectively. Taking into account the adsorbent doses used in the study, SW800 was superior to SW500. At the initial contact time, the rapid increase in adsorption was explained by the excess of vacant areas on the adsorbent surface, and as the sorption continues, the adsorption rate decreases with the decrease of the active areas on the sorbent surface.[3,14]

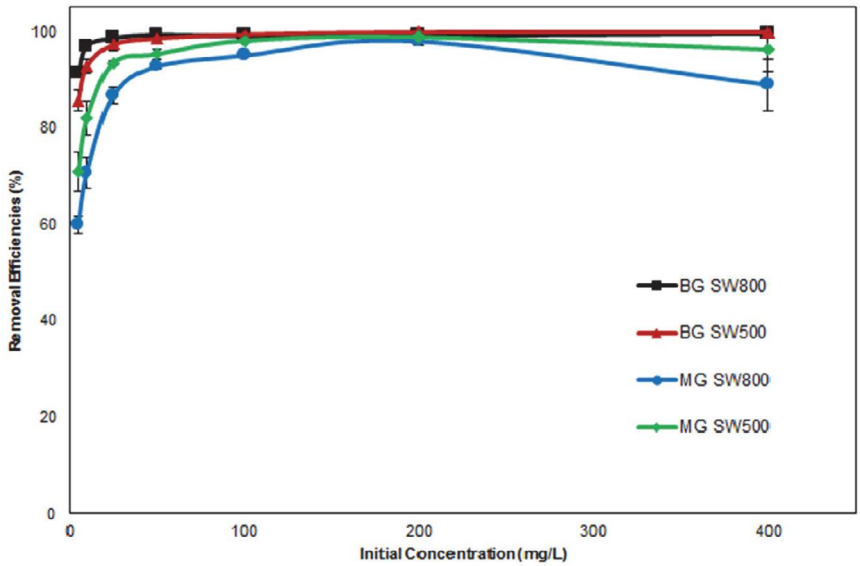


Figure 7. Effect of initial concentration on the removal of BG and MG (adsorbent dose of BG and MG: 0.4 and 0.6 g/L for SW800 and, 1.2 and 2.4 g/L for SW500, initial dye concentration range 5–400 mg/L, volume 25 mL, contact time 24 h, n = 3)

3. 2. 4. Effect of Initial Concentration on BG and MG Removal

The variations in removal efficiencies were investigated with initial dye concentrations ranging from 5 to 400 mg/L. The results are depicted in Figure 7. Sudden sharp increases were observed at the lower concentrations of BG and MG for both the adsorbents. The removal efficiencies became constant and then decreased at higher concentrations. It is thought that there is fixed number of available sites per unit mass of the adsorbent on the adsorbent surface. The number of available sites is higher for low initial dye concentration as against to the high initial concentration. Consequently, the most of the dye molecules are adsorbed by adsorbents at low initial dye concentrations and the removal efficiencies increases. On the other hand, when the certain initial dye concentrations is exceed, active sites of the adsorbent are completely retained, some of the dye molecules cannot be adsorbed, and the removal efficiencies become to decrease.[2,35] The removal efficiencies of BG reached equilibrium by 100 mg/L with 99.4% and 50 mg/L with 99.3% for SW500 and SW800, respectively. The removal efficiencies of MG reached equilibrium by 100 mg/L with 98.1% and 100 mg/L with 95.5% for SW500 and SW800, respectively.

3. 2. 5. Thermodynamic Studies

The effect of the temperature on BG and MG removal was investigated at 298, 303, 313 and 323 K. In order to calculate the thermodynamic parameters associated with the adsorption process, a change in Gibb's free energy (ΔG°), enthalpy (ΔH°) and entropy changes (ΔS°), used the equations below. The parameters exhibit spontaneity, randomness, and endothermicity/exothermicity of the adsorption processes.

$$\ln K_L = -\frac{\Delta H^o}{RT} + \frac{\Delta S^o}{R} \quad (3)$$

$$\Delta G^0 = -RT\ln K_L \quad (4)$$

$$\Delta G^o = \Delta H^o - T\Delta S^o \quad (5)$$

ΔG° is the free energy change (kJ/mol), R is the gas constant (8.314 J/mol K), K_L is the Langmuir equilibrium constant (L/mol),[36] and T is the temperature (K). K_L values were found from the ratio of the adsorbed dye concentration (mg) and equilibrium dye concentration in the solution (mg/L). The parameters of ΔH and ΔS were obtained from the slope and the intercept of the Van't Hoff graph between $\ln K_L$ and 1/T, respectively.[14]

The negative value of ΔG^o indicates the adsorption is spontaneous and favorable. ΔH^o values are positive whether the adsorption is endothermic, or vice versa. The positive ΔS^o reveals that randomness increased at the solid-liquid interface. The positive ΔS° also indicates the affinity of the adsorbent for BG and MG.[37,38] The calculated thermodynamic parameters are depicted in Table 1. As seen in Table 1, the adsorptions of BG and MG by both adsorbents were endothermic, favorable, and spontaneous. It was understood that the degree of randomness increased during the adsorption (ΔS° > 0). The value of ΔH^o presents an idea about different physical forces being involved in the adsorption process such as van der Waals forces (4–10 kJ/mol), hydrophobic bond forces (5 kJ/mol), hydrogen bond forces (2–40 kJ/mol), coordination exchange (40 kJ/mol), dipole bond forces (2–29 kJ/mol), and for chemical forces (>60 kJ/mol).[39] Our results indicated that the forces affecting the adsorption of BG could be hydrogen bond forces and dipole bond forces because of ΔH° values belonging to BG being calculated in our study as 2.65 and 28.50 kJ/mol for SW800 and SW500, respectively. However, ΔH° values of MG were calculated to be 28.38 and 47.79 kJ/mol for SW800 and SW500, respectively. Hence, it was thought that the adsorption of MG could be affected by hydrogen bond forces.

Table 1. Thermodynamic parameters for adsorption of BG and MG onto SW500 and SW800 (n = 3).

	T (K)	ΔH° (kJ/mol)	ΔS° (J/mol.K)	ΔG° (kJ/mol)
Brilliant Green				
SW800	298	2.650	103.9	–13.10
	303			–13.36
	313			–13.90
	323			–14.42
SW500	298	28.50	176.1	–8.972
	303			–9.372
	313			–10.17
	323			–10.97
Malachite Green				
SW800	298	28.38	174.3	–8.361
	303			–8.977
	313			–10.21
	323			–11.44
SW500w	298	47.79	219.9	–17.74
	303			–18.84
	313			–21.04
	323			–23.24

3. 2. 6. Isotherm Studies

The isotherm studies display the way of interactions between the dye molecules and the adsorbent, and also provide information about the nature of interactions. Experimental data was applied to the Langmuir,[40] Freundlich and Dubinin Radushkevic (D-R) isotherm models.[41] The Langmuir isotherm is based on the acceptance that the adsorption occurred at specific homogenous sites within the adsorbent while the Freundlich isotherm mentions the acceptance of a heterogeneous surface with a non-uniform distribution of heat of adsorption over the surface.[38] D-R

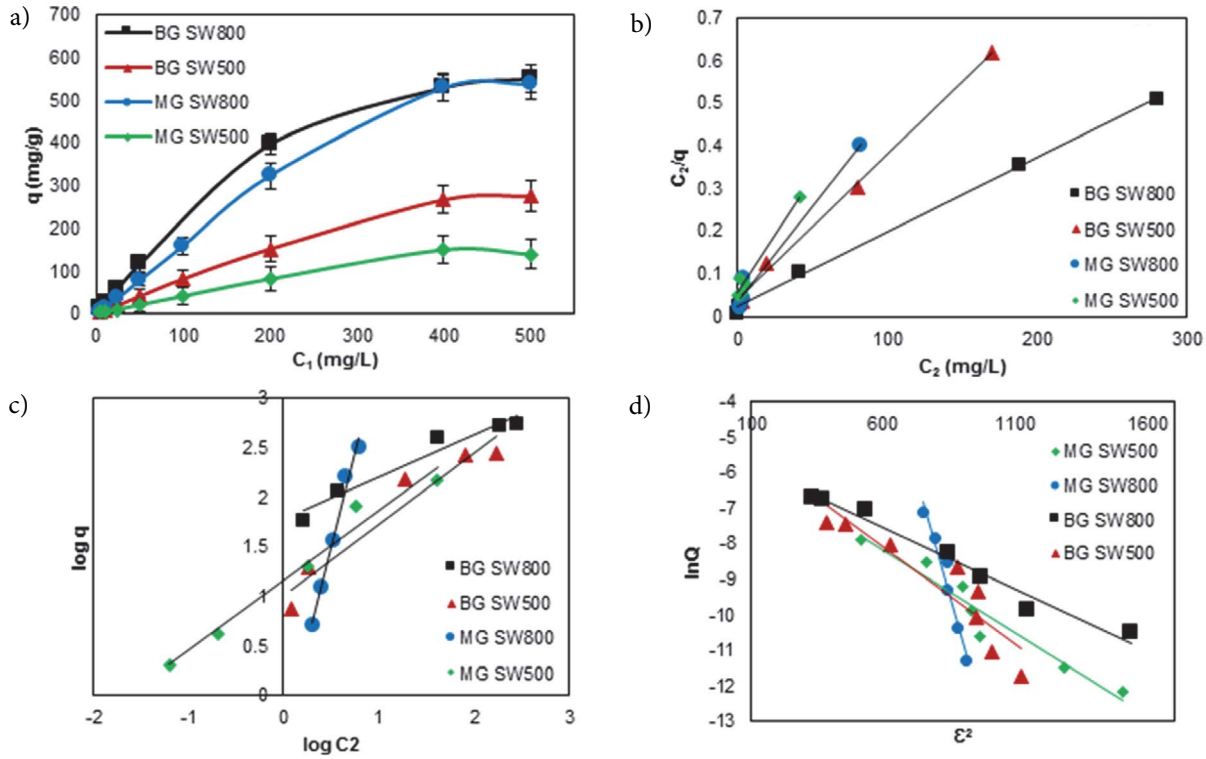


Figure 8. Adsorbed amount of dye as a function of initial concentration (a), Langmuir isotherms (b), Freundlich isotherms (c) and DR isotherms (d) for adsorption BG and MG, (initial dye concentration range 5–500 mg/L, volume 25 mL, adsorbent dose of BG and MG: 0.6 and 0.4 g/L for SW800 and, 1.2 and 2.4 g/L for SW500, contact time 24 h, n = 3)

Table 2. The isotherm parameters of Langmuir, Freundlich, and DR isotherms for BG and MG adsorption using SW500 and SW800 (n = 3).

		Brilliant Green		Malachite Green	
		SW800	SW500	SW800	SW500
Langmuir Isotherm					
$\frac{C_2}{q} = \left(\frac{1}{Q_{max}}\right) C_2 + \frac{1}{bQ_{max}}$	Q_{max}(mg/g)	555.6	294.1	222.2	185.19
	b (L/mg)	0.0833	0.0854	0.1372	0.1013
	R^2	0.9981	0.9963	0.9739	0.9732
	Separation factor	0.02–0.71	0.02–0.07	0.014–0.59	0.019–0.66
Freundlich Isotherm					
$\log q = \log K + \frac{1}{n} \log C_2$	1/n	0.4271	0.7233	3.7805	0.7041
	K (mg/g)(L/mg)$^{1/n}$	59.01	10.15	0.3975	14.3
	R^2	0.9541	0.9289	0.9882	0.9746
D-R Isotherm					
$\ln Q = \ln Q_m - k\varepsilon^2$	E (kJ/mol)	11.95	9.535	10.43	4.360
	Q_m (mol/g)	0.0041	0.0079	0.0044	4.251×10^6
$E = (2k)^{-0.5}$	k (mol^2/kJ^2)	0.0035	0.0055	0.0046	0.0263
	R^2	0.9741	0.8421	0.9351	0.95

C_2 is the equilibrium concentration of the solution (mg/L), q is the amount of adsorbed dye/amount of adsorbent (mg/g), b is the Langmuir constant (L/mg), Q_{max} is the monolayer adsorption capacity (mg/g), K is the Freundlich constant ((mg/g)(L/mg)$^{1/n}$), and 1/n is a dimensionless Freundlich constant for the intensity of the adsorbent, ε **(Polanyi potential) is (RTln(1 + 1/C_2))**, Q is the amount of dye adsorbed per unit weight of adsorbent (mol/g), Q_m is the adsorption capacity (mol/g), k is a constant related to adsorption energy (mol^2/kJ^2), R is the gas constant (kJ/mol K), and T is the absolute temperature (K).

isotherm expresses the mechanism of adsorption onto a heterogeneous surface.[42] In order to evaluate the adsorption isotherm, the used parameters were 25 mL of volume, an adsorbent dose of BG and MG 0.4 and 0.6 g/L for SW800 and 1.2 and 2.4 g/L for SW500, respectively and 24 h of contact time. Initial concentrations were in the range of 5–500 mg/L. Results and related equations were presented in Table 2 and Figure 8.

The correlation coefficients were evaluated to find the best fit isotherm model for the system. As seen in Table

2, the highest correlation coefficient (R^2) of BG was obtained for the Langmuir isotherm model while the highest R^2 of MG was for Freundlich isotherm model. Thus, the adsorption of BG by SW500 and SW800 were monolayer on homogeneous sites. However, the adsorption of MG was a multilayer adsorption on a heterogeneous site. The maximum monolayer adsorption capacities of BG and MG were calculated to be 294.1 mg/g and 185.19 mg/g for SW500 and 555.6 mg/g and 222.3 mg/g for SW800. 1/n values indicate the adsorption intensity. The higher 1/n values mean the higher affinity between the dye molecules and adsorbent.[38] The separation factor (R_L) shows whether the adsorption is favorable ($0 < R_L < 1$), unfavorable ($R_L > 1$), linear ($R_L = 1$) or irreversible ($R_L = 0$).[43] The separation factor is calculated by the equations given:

$$R_L = \left(\frac{1}{1+bC_1}\right) \quad (6)$$

where C_1 is the initial concentration and b is the Langmuir isotherm constant. Seeing that the R_L values were in the range of 0–1, adsorption was favorable for both the adsorbents.

The mechanism of adsorption can be determined by assessing E value (kJ/mol). The mean free energy of adsorption (E), which is defined as the free energy change when one mole of ion is transferred to the surface of a solid from the infinite space in the solution. Physical adsorption is valid if the value is below 8 kJ/mol. When the E value is between 8 kJ/mol and 16 kJ/mol, chemisorption or ion exchange occurs.[44] Since the values of BG (11.95; 9.54 kJ/mol for SW800; SW500) and MG (10.43; 4.36 kJ/mol for SW800; SW500) were between 8 and 16 kJ/mol, the presence of chemisorption or ion exchange could be mentioned.

3. 2. 7. Adsorption Kinetic Studies

In this study, to understand the adsorption mechanism, three simplified kinetic models were elucidated: Lagergren pseudo first order, pseudo second order and intraparticle diffusion model.[41] These models define the stages of the adsorption to be external film diffusion, adsorption, and intraparticle diffusion.[45] Figure 9 presents the time effects on the adsorption, pseudo first-order kinetic model, pseudo second-order kinetic model, and intraparticle diffusion kinetic model for the adsorption of BG and MG onto SW500 and SW800 adsorbents. The calculated parameters belonging to the kinetic models are depicted in Table 3. As can be seen from Table 3, the closest R^2 values to unity were obtained for the pseudo second order kinetic model for the studied dyes. The calculated ($q_{e,\,cal}$) and experimental ($q_{e,exp}$) values of adsorption capacities of BG and MG were very close to each other for the pseudo second order kinetic model. These findings indicated the adsorption fitted well with pseudo second order kinetic model and adsorption was chemisorption controlled.[46] In the

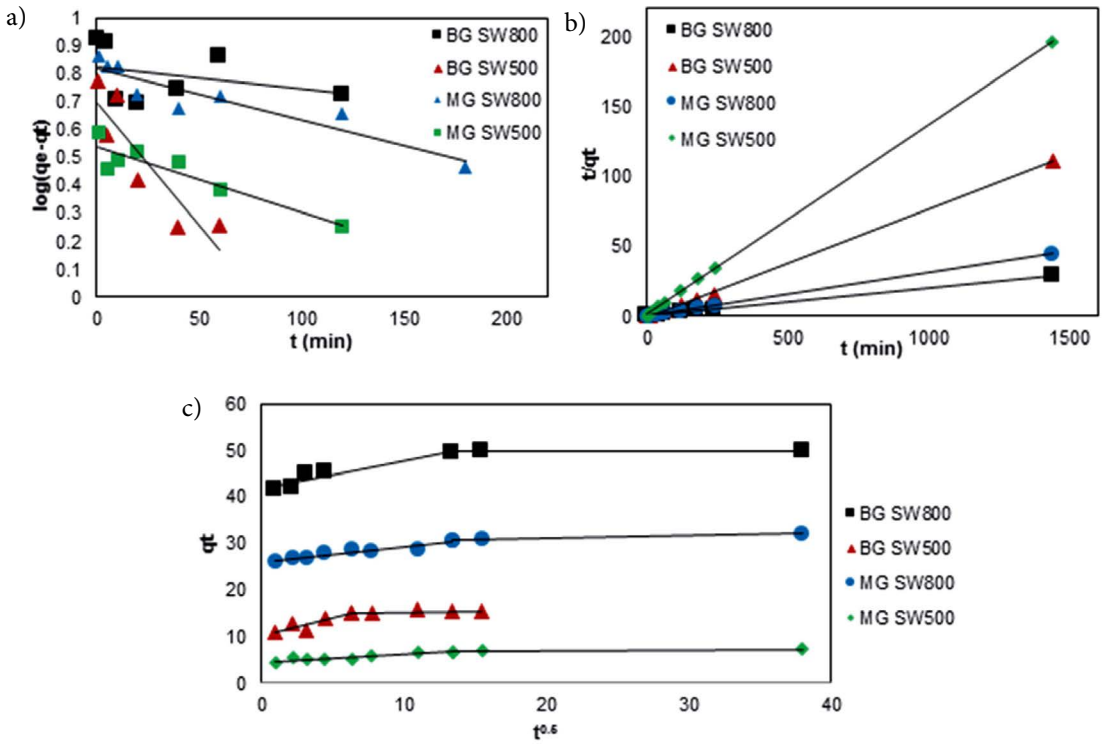


Figure 9. Kinetic studies for BG and MG adsorption a) pseudo first-order kinetic model, b) pseudo second-order kinetic model, c) intraparticle diffusion kinetic model (initial dye concentration 20 mg/L, volume 25 mL, adsorbent dose of BG and MG: 0.4 and 0.6 g/L for SW800 and, 1.2 and 2.4 g/L for SW500, contact time 24 h, temperature 25 °C, n = 3)

Table 3. The constants of the pseudo first-order, pseudo second-order kinetic models, and intraparticle kinetic model for BG and MG removal (n = 3).

Kinetic Models	Adsorbent SW800				SW500			
	$q_{e,exp}$ (mg/g)	$q_{e,cal}$ (mg/g)	k_1 (1/min)	R^2	$q_{e,exp}$ (mg/g)	$q_{e,cal}$ (mg/g)	k_2 (g/mgmin)	R^2
Pseudo first order								
$\log(q_e - q_t) = \log q_e - \frac{k_1 t}{2.303}$								
BG	49.67	6.668	0.00184	0.1121	14.84	5.041	0.0205	0.7996
MG	33.33	6.637	0.00438	0.8687	8.33	3.472	0.0053	0.8469
Pseudo second order								
$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{t}{q_e}$								
BG	49.67	50	0.005076	0.9999	14.84	12.89	–0.00735	0.9990
MG	33.33	32.26	0.005128	0.9999	8.33	7.369	0.01363	0.998
Intraparticle diffusion model								
$q_t = k_{int} t^{0.5} + I$		I (mg/g)	k (mg/gmin$^{0.5}$)	R^2		I (mg/g)	k (mg/gmin$^{0.5}$)	R^2
BG		Step 1	41.63	0.6047	0.8893	10.19	0.7498	0.7789
		Step 2	49.483	0.0082	0.2363	14.522	0.0647	0.5396
MG		Step 1	25.967	0.3118	0.8969	4.5719	0.1621	0.8509
		Step 2	29.652	0.0654	0.9851	6.5568	0.0207	0.7878

q*e* and qt indicate the adsorption capacity at equilibrium (mg/g) and at time t; k1 and k2 are the pseudo first-order (1/min) and pseudo second-order rate constants (g/mg min); t is the contact time (min) and kint (mg/g min$^{0.5}$) and I (mg/g) are the intraparticle diffusion constants.

intraparticle diffusion model, the plot q_t versus $t^{0.5}$ gives k and I as slope and intercept, respectively. The intercept indicates the effect of the boundary layer thickness. The higher the intercept length, the more the adsorption is boundary layer controlled.[46] Also, if the line passes through the origin (I = 0), the rate limiting mechanism is solely controlled by the intraparticle diffusion. Thus, it was concluded that the intraparticle diffusion was not the only rate limiting step. Since two separate regions were obtained for both the adsorbents, the adsorption process was affected by two or more steps. The initial region is ascribed to the bulk diffusion while the second to the intraparticle diffusion.[38]

4. Conclusion

The adsorbents used in this study were obtained from the sandpaper wastes. Hazardous and toxic reagents were not used during the preparation of the adsorbents. Thus, environmentally friendly adsorbents were obtained. Furthermore, the removal of brilliant green from aqueous solutions was successfully carried out using both the adsorbents. Optimization studies (pH, adsorbent dose, contact time, and initial concentration etc.) were carried out to investigate the removal performance of both adsorbents. According to the pH study, SW800 provided a wider pH range than that of SW500 for both dyes. The optimum adsorbent doses of BG and MG were selected as 0.4 and 0.6 g/L for SW800 while 1.2 and 2.4 g/L for SW500, respectively. The removal efficiencies of BG and MG reached a plateau after 120 min and 180 min for SW500 and 180 min and 240 min for SW800, respectively. The adsorption kinetics of the dyes fitted well with the pseudo second-order kinetic model. The adsorption of the BG showed good agreement with the Langmuir isotherm model and indicated monolayer adsorption on homogeneous sites. However, it was found that the adsorption of the MG obeyed the Freundlich isotherm model. The values of E indicated the adsorption mechanism of dyes could be chemical or through ion exchange. The adsorption of BG and MG were found to be favorable for both SW500 and SW800. The thermodynamic studies indicated that the process was endothermic, spontaneous, and feasible. The comparison of the maximum BG and MG adsorption capacities with the reported adsorbents in the literature can be found in Table 4. By comparing the maximum adsorption capacities in Table 4, the highest capacity values belonged to the sandpaper waste. As a result, environmentally friendly adsorbents were developed that facilitate fast and efficient removal.

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Table 4 Comparison of the maximum adsorption capacities of BG and MG with the reported adsorbents in the literature.

BG					MG				
Adsorbent	**Capacity (mg/g)**	**Isotherm**	**Kinetic model**	**Ref.**	**Adsorbent**	**Capacity (mg/g)**	**Isotherm**	**Kinetic model**	**Ref.**
Tannin gel (TG)	8.55	Langmuir	Pseudo second order	6	AMP clay	130.64	Langmuir	Pseudo first order	1
Amine modified TG	2.41	Langmuir	Pseudo second order	6	Coconot coir activated carbon	27.44	Langmuir Freundlich	Pseudo second order	7
Acorn	2.01	Langmuir	Pseudo second order	46	Rattan sawdust	62.71	Langmuir	Pseudo first order	12
Peganum harmala-L seeds	35.97	Langmuir	Pseudo second order	3	EM based compost	159.22	Sips	Pseudo second order	13
Saklıkent mud	1.18	Langmuir	Pseudo second order	45	Potato peel	35.61	Redlich-Peterson	Pseudo-nth order	50
White rice husk ash	85.56	–	Pseudo second order	16	GO–Fe_3O_4	160.7	Langmuir Freundlich	Pseudo second order	51
Red clay	125	Redlich-Peterson	Pseudo second order	35	Citrus limetta peel	8.733	D-R	Pseudo second order	22
Kaolin	65.42	Langmuir	Pseudo second order	2	Zea mays cob	16.72	D-R	Pseudo second order	22
NaOH treated saw dust	58.48	Redlich-Peterson and Temkin	Pseudo second order	14	Organically modified hydroxyapatite	188.18	Langmuir	–	21
Ni/Ni_xB nanoparticle-coated resin	147.1	Langmuir	Pseudo second order	49	SW800	222.2		Pseudo second order	This study
SW800	555.6	Langmuir	Pseudo second order	This study	SW500	185.19		Pseudo second order	This study
SW500	294.1	Langmuir	Pseudo second order	This study					

5. References

1. Y.-C. Lee, E. J. Kim, J.-W. Yang and H.-J. Shin, *J. Hazard. Mater.* **2011**, *192*, 62–70.
2. B. K. Nandi, A. Goswami and M. K. Purkait, *J. Hazard. Mater.* **2009**, *161*, 387–395. **DOI:**10.1016/j.jhazmat.2008.03.110
3. S. Agarwal, V. K. Gupta, M. Ghasemi and J. Azimi-Amin, *J. Mol. Liq.* **2017**, *231*, 296–305. **DOI:**10.1016/j.molliq.2017.01.097
4. L. Kong, F. Qiu, Z. Zhao, X. Zhang, T. Zhang, J. Pan and D. Yang, *J. Clean. Prod.* **2016**, *137*, 51–59. **DOI:**10.1016/j.jclepro.2016.07.067
5. M. Oplatowska, R. F. Donnelly, R. J. Majithiya, D. Glenn Kennedy and C. T. Elliott, *Food Chem. Toxicol.* **2011**, *49*, 1870–1876. **DOI:**10.1016/j.fct.2011.05.005
6. N. Akter, A. Hossain, M. J. Hassan, M. K. Amin, M. Elias, M. M. Rahman, A. M. Asiri, I. A. Siddiquey and M. A. Hasnat, *J. Environ. Chem. Eng.* **2016**, *4*, 1231–1241. **DOI:**10.1016/j.jece.2016.01.013
7. Uma, S. Banerjee and Y. C. Sharma, *J. Ind. Eng. Chem.* **2013**, *19*, 1099–1105. **DOI:**10.1016/j.jiec.2012.11.030
8. A. A. Adeyemo, I. O. Adeoye and O. S. Bello, *Appl. Water Sci.* **2017**, *7*, 543–568. **DOI:**10.1007/s13201-015-0322-y
9. L. Bulgariu, L. B. Escudero, O. S. Bello, M. Iqbal, J. Nisar, K. A. Adegoke, F. Alakhras, M. Kornaros and I. Anastopoulos, *J. Mol. Liq.* **2019**, *276*, 728–747. **DOI:**10.1016/j.molliq.2018.12.001
10. A. Kausar, M. Iqbal, A. Javed, K. Aftab, Z.-H. Nazli, H. N. Bhatti and S. Nouren, *J. Mol. Liq.* **2018**, *256*, 395–407. **DOI:**10.1016/j.molliq.2018.02.034
11. V. Katheresan, J. Kansedo and S. Y. Lau, *J. Environ. Chem. Eng.* **2018**, *6*, 4676–4697. **DOI:**10.1016/j.jece.2018.06.060
12. B. H. Hameed and M. I. El-Khaiary, *J. Hazard. Mater.* **2008**, *159*, 574–579. **DOI:**10.1016/j.jhazmat.2008.02.054
13. T. Bhagavathi Pushpa, J. Vijayaraghavan, S. J. Sardhar Basha, V. Sekaran, K. Vijayaraghavan and J. Jegan, *Ecotoxicol. Environ. Saf.* **2015**, *118*, 177–182. **DOI:**10.1016/j.ecoenv.2015.04.033
14. V. S. Mane and P. V. V. Babu, *Desalination* **2011**, *273*, 321–329. **DOI:**10.1016/j.desal.2011.01.049
15. V. K. Gupta, B. Gupta, A. Rastogi, S. Agarwal and A. Nayak, *J. Hazard. Mater.* **2011**, *186*, 891–901. **DOI:**10.1016/j.jhazmat.2010.11.091
16. M. P. Tavlieva, S. D. Genieva, V. G. Georgieva and L. T. Vlaev, *J. Colloid Interface Sci.* **2013**, *409*, 112–122. **DOI:**10.1016/j.jcis.2013.07.052

17. K. G. Bhattacharyya and A. Sarma, *Dye. Pigment.* **2003**, *57*, 211–222. **DOI:**10.1016/S0143-7208(03)00009-3
18. T. Uysal, G. Duman, Y. Onal, I. Yasa and J. Yanik, *J. Anal. Appl. Pyrolysis* **2014**, *108*, 47–55. **DOI:**10.1016/j.jaap.2014.05.017
19. M. Baghdadi, B. A. Soltani and M. Nourani, *J. Ind. Eng. Chem.* **2017**, *55*, 128–139. **DOI:**10.1016/j.jiec.2017.06.037
20. Sandpaper, https://en.wikipedia.org/wiki/Sandpaper, (accessed 1 November **2018**).
21. A. A. El-Zahhar and N. S. Awwad, *J. Environ. Chem. Eng.* **2016**, *4*, 633–638. **DOI:**10.1016/j.jece.2015.12.014
22. H. Singh, G. Chauhan, A. K. Jain and S. K. Sharma, *J. Environ. Chem. Eng.* **2017**, *5*, 122–135. **DOI:**10.1016/j.jece.2016.11.030
23. S. Milicevic, T. Boljanac, S. Martinovic, M. Vlahovic, V. Milosevic and B. Babic, *Fuel Process. Technol.* **2012**, *95*, 1–7. **DOI:**10.1016/j.fuproc.2011.11.005
24. M. J. Rwiza, S. Y. Oh, K. W. Kim and S. D. Kim, *Chemosphere* **2018**, *195*, 135–145. **DOI:**10.1016/j.chemosphere.2017.12.043
25. N. Kataria and V. K. Garg, *J. Environ. Chem. Eng.* **2017**, *5*, 5420–5428. **DOI:**10.1016/j.jece.2017.10.035
26. C. L. Lu, J. G. Lv, L. Xu, X. F. Guo, W. H. Hou, Y. Hu and H. Huang, *Nanotechnology* **2009**, *20*, 215604–215612. **DOI:**10.1088/0957-4484/20/21/215604
27. H. Xia, L. Chen and Y. Fang, *Sep. Sci. Technol.* **2013**, *48*, 2681–2687. **DOI:**10.1080/01496395.2013.805340
28. L. Dai, W. Zhu, L. He, F. Tan, N. Zhu, Q. Zhou, M. He and G. Hu, *Bioresour. Technol.* **2018**, *267*, 510–516. **DOI:**10.1016/j.biortech.2018.07.090
29. K.-W. Jung, B. H. Choi, M.-J. Hwang, T.-U. Jeong and K.-H. Ahn, *Bioresour. Technol.* **2016**, *219*, 185–195. **DOI:**10.1016/j.biortech.2016.07.098
30. S. Chowdhury and P. Saha, *Chem. Eng. J.* **2010**, *164*, 168–177. **DOI:**10.1016/j.cej.2010.08.050
31. I. A. Aguayo-Villarreal, L. A. Ramírez-Montoya, V. Hernández-Montoya, A. Bonilla-Petriciolet, M. A. Montes-Morán and E. M. Ramírez-López, *Ind. Crops Prod.* **2013**, *48*, 89–97. **DOI:**10.1016/j.indcrop.2013.04.009
32. N. Fiol and I. Villaescusa, *Environ. Chem. Lett.* **2009**, *7*, 79–84. **DOI:**10.1007/s10311-008-0139-0
33. F. de Castro Silva, M. M. F. da Silva, L. C. B. Lima, J. A. Osajima and E. C. da Silva Filho, *Int. J. Biol. Macromol.* **2018**, *114*, 470–478. **DOI:**10.1016/j.ijbiomac.2018.03.089
34. A. B. Karim, B. Mounir, M. Hachkar, M. Bakasse and A. Yaacoubi, *J. Hazard. Mater.* **2009**, *168*, 304–309. **DOI:**10.1016/j.jhazmat.2009.02.028
35. M. Saif, U. Rehman, M. Munir, M. Ashfaq, M. F. Nazar, M. Danish and J. Han, *Chem. Eng. J.* **2013**, *228*, 54–62. **DOI:**10.1016/j.cej.2013.04.094
36. E. C. Lima, A. Hosseini-Bandegharaei, J. C. Moreno-Piraján and I. Anastopoulos, *J. Mol. Liq.* **2019**, *273*, 425–434. **DOI:**10.1016/j.molliq.2018.10.048
37. Y. Bulut and H. Aydin, *Desalination* **2006**, *194*, 259–267. **DOI:**10.1016/j.desal.2005.10.032
38. V. S. Mane, I. D. Mall and V. C. Srivastava, *J. Environ. Manag.* **2007**, *84*, 390–400. **DOI:**10.1016/j.jenvman.2006.06.024
39. S. K. Srivastava, V. K. Gupta, M. K. Dwivedi and S. Jain, *Anal. Proc. Incl. Anal. Commun.* **1995**, *32*, 21–23. **DOI:**10.1039/AI9953200021
40. I. Langmuir, *J. Am. Chem. Soc.* **1918**, 40, 1361–1403. **DOI:**10.1021/ja02242a004
41. T. D. Çiftçi, *Cogent Chem.* **2017**, *3*, 1–15. **DOI:**10.1080/23312009.2017.1284296
42. M. Ghasemi, M. Naushad, N. Ghasemi and Y. Khosravi-fard, *J. Ind. Eng. Chem.* **2014**, *20*, 2193–2199. **DOI:**10.1016/j.jiec.2013.09.050
43. A. K. Meena, K. Kadirvelu, G. K. Mishraa, C. Rajagopal and P. N. Nagar, *J. Hazard. Mater.* **2008**, *150*, 619–625. **DOI:**10.1016/j.jhazmat.2007.05.011
44. N. K. Amin, *J. Hazard. Mater.* **2009**, *165*, 52–62. **DOI:**10.1016/j.jhazmat.2008.09.067
45. Y. Kismir and A. Z. Aroguz, *Chem. Eng. J.* **2011**, *172*, 199–206. **DOI:**10.1016/j.cej.2011.05.090
46. M. Ghaedi, H. Hossainian, M. Montazerozohori, A. Shokrollahi, F. Shojaipour, M. Soylak and M. K. Purkait, *Desalination* **2011**, *281*, 226–233. **DOI:**10.1016/j.desal.2011.07.068
47. N. Akter, M. A. Hossain, M. J. Hassan, M. K. Amin, M. Elias, M. M. Rahman, A. M. Asiri, I. A. Siddiquey and M. A. Hasnat, *J. Environ. Chem. Eng.* **2016**, *4*, 1231–1241. **DOI:**10.1016/j.jece.2016.01.013
48. M. S. Raghu, K. Y. Kumar, M. K. Prashanth, B. P. Prasanna, R. Vinuth and C. B. Pradeep Kumar, *J. Water Process Eng.* **2017**, *17*, 22–31. **DOI:**10.1016/j.jwpe.2017.03.001
49. M. Çınar, Y. İşlek Coşkun and T. Deniz Çiftçi, *Turkish J. Chem.* **2018**, *42*, 505–519.
50. E-K. Guechi and O. Hamdaoui, Arab. J. Chem. **2016**, *9*, 416–424. **DOI:**10.1016/j.arabjc.2011.05.011
51. M. S. Raghu, K.Y. Kumar, M.K. Prashanth et al., J. Water Process Eng. **2017**, *17*, 22–31. **DOI:**10.1016/j.jwpe.2017.03.001

Povzetek

Odpadke brusilnega papirja smo po pirolizi na 500 in kalcinaciji na 600 uporabili za odstranjevanje kationskih barvil briljantno zeleno in malahitno zeleno iz vodnih raztopin. Adsorpcijsko ravnotežje smo poskusili opisati z Langmuir-jevo, Freundlich-ovo, in Dubinin-Radushkevich-ovo izotermo ter izvedli termodinamske študije. Adsorbcijo na briljantno zeleno smo najbolje opisali z Langmuir-jevo izotermo, adsorpcijo na malahitno zeleno pa z Freundlich-ovo. Izkazalo se je, da je adsorpcija endotermna. E vrednost pridobljena iz Dubinin-Radushkevich-ove izoterme je pokazala, da gre za kemijsko adsorpcijo. Kinetiko adsorpcije smo preučili s tremi kinetični modeli (psevdo prvi-red, psevdo drugi-red reakcije in modelom znotraj-delčne difuzije) in pokazali, da model psevdo–drugega reda najbolje opiše adsorpcijo na obe barvili.