Scientific paper

Exploring Bikaverin as Metal Ion Biosensor: A Computational Approach

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Abstract

A computational exploration of fungi produced pigment bikaverin as a biosensor towards bioavailable metal ions is presented. Systematic studies of the optimized ground and excited state geometries were attempted for exploring metal ion binding pocket, comparative binding propensity and optical properties of the bikaverin and its adducts with studied metal ions. The screening of thirteen (13) bioavailable metal ions, revealed a range of binding strength towards bikaverin receptor with Ca^{2+} , Mg^{2+} and Al^{3+} as the strongest binders. Besides, upon binding to bikaverin receptor an enhancement in its fluorescence intensity was observed in the order $Ca^{2+} > Al^{3+} > Mg^{2+}$. The computationally predicted selectivity of bikaverin receptor towards Ca^{2+} was experimentally corroborated through the preliminary fluorescence studies. The bikaverin probe showed an enhancement of fluorescence emission in presence of Ca^{2+} ions in buffered aqueous medium.

Keywords: Computational chemistry; Biosensors; Fluorescence Spectroscopy; Quantum chemical calculations; Electronic structure

1. Introduction

The well designed computational studies can be good to predict the starting point for targeted experiments in a time and cost effective manner. Experimental chemists use theoretical calculations to supplement, support and guide their experiments particularly in the areas of conformational analysis,² reaction mechanism,³ transition states,4 charge distributions,5 modeling larger molecules like DNA and proteins,6 structure-activity relationships,7 orbital interactions,8 and excited state studies.9 Metal ions are required for a plethora of functions in biosystems. However, the metal ions can be like double-edged swords; at their proper concentrations, these remain coordinated to their natural binding sites and perform the desired functions; but a change in their normal concentration can lead to their de-compartmentalization and consequently bring onset of deleterious functions. 10 Therefore, selective sensors for detection and quantification of metal ions for the real time analytical monitoring and medical diagnosis are of considerable importance.¹¹

Thus, it becomes imperative to investigate the natural bioactive compounds for their in vivo metal ion sensing applications in order to potentiate the diagnosis for better therapeutic action. In continuation of our computational chemistry interests^{12–15} and towards investigating the bioactivities of natural products, 16-18 we attempted to survey the fungi derived fluorescent bioactive natural product bikaverin for metal ion sensing ability. Bikaverin is a reddish pigment produced by different fungal species of the genus Fusarium with the diverse biological activities. 19-21 With the increasing reports on its pharmacological role, bikaverin is becoming a metabolite of biotechnological interest.²² Commercially bikaverin as pure compound is costly and synthesizing bikaverin through a total chemical process is less viable and environmentally unfavorable. However, obtaining bikaverin from microbial sources present an environment friendly, continuous and cheap source. Thus for obtaining bikaverin in a cost effective and an environment friendly manner, we extracted bikaverin from fungal extracts using the standardized procedure from literature reports.²³ In this study, we present the computational exploration of bikaverin as biosensor towards thirteen (13) bioavailable metal ions. The computational prediction was experimentally confirmed through preliminary florescence studies of studied metal ions as bikaverin adducts.

2. Experimental

2. 1. Computational Procedures and Materials

All density functional theory (DFT) and time-dependent density functional theory (TD-DFT) calculations were performed using the Gaussian 09 program package.²⁴ The geometries of all the studied molecules (bikaverin and its metals adducts) were fully optimized at the singlet ground (S0) and first excited (S1) states in the acetonitrile solvent using CAM-B3LYP, Coulomb-attenuating method based on Becke's three parameter hybrid exchange and nonlocal correlation functional of Lee, Yang and Parr (65% exchange and 35% correlation weighting at longrange). 25,26 The standard 6-311G, split-valence atomic basis set function of DFT has been employed for all the atoms except for the transition metal ions; Nickel (Ni²⁺), Mercury (Hg²⁺), Cadmium (Cd²⁺), Manganese(Mn²⁺), Iron (Fe²⁺), Zinc(Zn²⁺), Copper(Cu²⁺) and Lead (Pb²⁺) atoms for which effective core potential (ECP) of Wadt and Hay pseudo-potential with a double-ζ valence basis set LANL2DZ was used.²⁷⁻²⁹ It is because the LANL2DZ reduces the computational cost as it uses an effective core potential for the core electrons, thus the core electrons are not explicitly considered in the computation. For the structural calculations of receptor-transition metal complexes, the relativistic LANL2DZ pseudopotential is the typical basis set asit has been identified to give close agreement between calculated and experimentally observed geometries. Frequency calculations were carried out to verify that the optimized molecular structures correspond to the energy minima, thus only positive frequencies were expected. The TD-DFT studies of all the molecules were performed by utilizing the same functional and the basis sets as CAM-B3LYP has been shown to be good for the excited state property calculations.³⁰ The Conductor-like Polarizable Continuum Model (CPCM) was utilized for taking care of the effect of solvation in acetonitrile (dielectric constant, $\varepsilon = 37.5$) on all the calculations as implemented in Gaussian 09. Natural Bond Orbital (NBO) method was employed for the natural population analysis, using the NBO 3.1 version as implemented in Gaussian 09 program.³¹ Free energy changes (ΔG) and binding energies (ΔE) were calculated for the receptor-analyte complexes to comprehend the thermodynamic binding propensity of the selected bioavailable cations with receptor (1). The Gibb's free energy change (ΔG) of the fragments (1), Na⁺, K⁺, Ni²⁺, Al³⁺, Hg²⁺, Cd²⁺, Mn²⁺, Fe²⁺, Zn²⁺, Cu²⁺, Mg²⁺, Pb²⁺ and Ca²⁺and their complexes were calculated from equation 1:

$$\Delta G = Gcomplex - (Greceptor + Gcation)$$
 (1)

Where Gcomplexis the free energy of receptor-cation adduct, Greceptor and Gcation are free energies of isolated receptor and cation respectively. The binding energy changes were also calculated to check the binding selectivity of the studied cations towards bikaverin receptor using the equation 2:

$$\Delta E = (Ecomplex) - (Ereceptor + Ecation)$$
 (2)

Where Ecomplex is the total energy of the receptorcation adduct, Ereceptor and Ecation are total energies of isolated receptor and cation respectively. In order to reduce basis set superposition error (BSSE) in these energy calculations, the Boys-Bernardi scheme was applied to yield the counterpoise corrected energies. ³²

2. 2. Extraction of Bikaverin from Fungal Extracts

Fungal mycelium showed a growth up to 5–6 cm on a PDA (potato dextrose agar) plate after 7 days. The morphological features supported the genus *Fusarium* resulting in 99% homology with the *Fusarium proliferatum*. Dichloromethane solvent was used to extract the crude bikaverin using National Cancer Institute's protocol. The extract was concentrated under vacuum and subjected to purification over column chromatography on silica gel using hexane-ethyl acetate (7:3 V:V) mobile phase leading to the isolation of pure bikaverin.

2. 3. Absorption and Emission Studies

All reagents were purchased from Aldrich and used without further purification. UV-Vis and fluorescence spectra were recorded on a Shimadzu UV-2450 and Shimadzu 5301 PC spectrophotometer respectively, with a quartz cuvette (path length 1 cm). A 10^{-3} M stock solution of bikaverin (probe 1) was prepared by dissolving the required amount of bikaverin in 10 mL of DMSO; 30µL of this stock solution was further diluted with CH3CN and HEPES buffer (0.05 M, pH = 7.4) to prepare 3 mL solution of probe (1) and this solution was used for each UV-Visible and fluorescence experiment. The aliquots of freshly prepared standard solutions (10⁻²M) of metal chloride/ metal perchlorates $\{M = Hg^{2+}, Fe^{2+}, Pb^{2+}, Cd^{2+}, Cu^{2+}, Cu^{$ Zn²⁺, Ni²⁺, Al³⁺, Co²⁺, Mg²⁺, Ca²⁺, Na⁺, and K⁺}, in distilled water were added to 3 mL solution of probe (1) taken in quartz cuvette and spectra's were recorded.

3. Results and Discussions

Quantum-chemical calculations are impressive for their step to step analysis and prediction whereas experi-

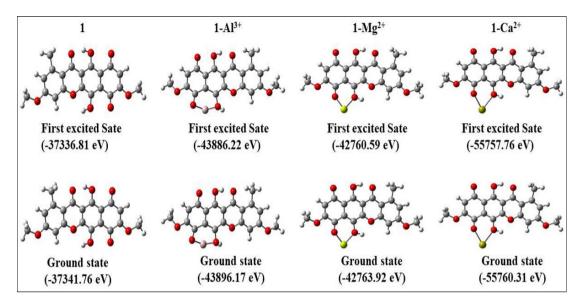


Figure 1. Optimized ground (S0) and the first excited state (S1) geometries of bikaverin receptor(1), 1-Al³⁺, 1-Mg²⁺ and 1-Ca²⁺ and their corresponding energies [in electron volts (eV)].

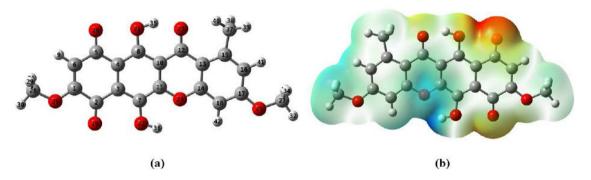


Figure 2. (a) Optimized geometry and (b) Electrostatic potential map of ground state (S₀) of bikaverin.

mental results are a net outcome of all the influencing factors in a blend. The optimized geometries and the corresponding energies of the ground and first excited state of

Table 1. The NPA Charge distribution on some crucial atoms of bikaverin 1 and $1\text{-}Ca^{2+}$ in S0 calculated at CAM-B3LYP/6-311G level of theory.

Atom involved	NPA charge distribution Bikaverin (1) 1-Ca ²⁺			
O22	-0.685	-0.776		
O19	-0.534	-0.678		
O23	-0.645	-0.635		
O21	-0.698	-0.685		
O20	-0.588	-0.559		
O24	-0.495	-0.495		
O25	-0.532	-0.527		
O26	-0.516	-0.513		
H36	-0.523	-0.551		
Ca43	••••	1.964		

bikaverin(1) and its receptor-analyte complexes with thirteen (13) studied bioavailable metal ions are shown in Figures 1, and S1 and S2 respectively (see Supplementary information). The natural charge distribution (Table 1) and electrostatic potential map of bikaverin (1) were calculated to identify its appropriate cation binding site. The charge distribution data in Table 1 and electrostatic potential map of bikaverin (1) indicate that the pocket for the electrophilic addition of cations is the carbonyl and the hydroxyl oxygen atoms Figure2 (see the ground state structure of bikaverin).

The calculated free energy change (ΔG) and the binding energy (ΔE) values of receptor analyte complexes are presented in Table 2. The data in Table 2 reveals that the binding of all the studied bioavailablecations to bikaverin is thermodynamically feasible and these ions bind the bikaverin receptor (1) with a range of strength. The trend in the binding propensity of studied metal ions can be correlated to their charge densities and hard soft acid base strength. It is evident from the binding and free energy data of Table 2 that the electrophillic binding of Ca^{2+} ,

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1-Zn2+

S. No	Adduct	ΔG (Kcal/mol)	Binding energy (Kcal/mol)	S. No	Adduct	ΔG (Kcal/mol)	Binding energy (Kcal/mol)
01	1-Na+	-22.44	-41.94	08	1-Mn ²⁺	-43.76	-31.81
02	1-K+	-19.92	-39.31	09	1-Fe ²⁺	-55.39	-50.11
03	1-Al ³⁺	-53.09	-52.96	10	1-Ni ²⁺	-27.11	-29.72
04	1-Pb ²⁺	-51.31	-22.72	11	1-Cu ²⁺	-34.64	-39.94
05	1-Hg ²⁺	-15.29	-34.20	12	$1-Mg^{2+}$	-51.97	-58.63
06	1-Cd ²⁺	-22.64	-45.29	13	1-Ca ²⁺	-73.19	-69.21

-23.77

Table 2. Calculated free energy change (ΔG) and binding energy for receptor-analyte complexes using CAM-B3LYP/6-311Glevel of theory with basis set superposition error (BSSE) corrections.

 ${\rm Mg^{2+}}$ and ${\rm Al^{3+}}$ with receptor (1) is stronger in comparison to the remaining cations. Among all the studied metal ions the binding of ${\rm Ca^{2+}}$ to bikaverin was found to be strongest which suggests a possible interaction between bikaverin and ${\rm Ca^{2+}}$ at lower concentrations. Thus a sensing application of bikaverin towards ${\rm Ca^{2+}}$ ions at its physiological concentrations can be possible

-31.66

Motivated by the binding of metal ions to bikaverin receptor(1), we attempted to explore the fluorescence behavior of the studied metal ions for predicted sensing applications. To get an insight into the origin of absorption and emission bands, the transitions of bikaverin(1) and its receptor-analyte complexes under consideration were simulated using preferred DFT/TD-DFT level of theory (Experimental section). The computationally predicted spectra of studied metal ion bikaverin (1) adducts in the acetonitrile solvent depicted various peaks. However only the peaks of good intensity fluorescence were chosen for analysis. The major absorption and fluorescence intensity band of bikaverin (1) and its receptor-analyte adducts with studied metal ions are summarized in Table 3. The calculated absorption spectrumfor bikaverin (1) shows a major peak (\lambda abs) at 491 nm (2.52 eV). The metal ion bikaverin adducts showed varied emission responses in which the fluorescence intensities in some cases were seen to be attenuated and in some cases enhanced. The calculated fluorescence intensities of 1-Ca²⁺, 1-Mg²⁺ and 1-Al³⁺adducts were found to be significantly higher than that of bikaverin (1) while as the absorption intensities of other receptor-analyte complexes don't show much deviation in comparison to pure bikaverin (1).

To have a further insight into the emission behavior of the bikaverin (1) metal ion adducts, the frontier molecular orbitals (FMOs) analysis was attempted. The FMOs of the bikaverin (1) and some representative receptor-analyte complexes are depicted in Fig. 3. The highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbitals (LUMO) energies of bikaverin (1) and all its metal ion adducts have been summarized in (Table S1, see supporting information). The interactions between the metal ions and the bikaverin (1) can be clearly observed from the frontier molecular orbital contour distribution. 33

Table 3. Summary of absorption and fluorescence spectral data of bikaverin(1) and its analyte complexes

Molecule	Calculated $\lambda_{abs}(nm)$	Oscillator strength (f)	Calculated $\lambda_{flu}(nm)$	Oscillator strength (f)
1	491	0.2524	576	0.0102
1-Na+	412	0.1960	426	0.0081
1-K+	315	0.1910	370	0.0013
1-Ni2+	441	0.2103	494	0.0014
1-Al3+	342	0.1543	388	0.1976
1-Hg2+	377	0.1130	435	0.0045
1-Cd2+	389	0.1866	437	0.0043
1-Mn2+	422	0.2010	497	0.0036
1-Fe2+	360	0.2349	454	0.0045
1-Zn2+	390	0.1873	437	0.0033
1-Cu2+	391	0.1033	463	0.0002
1-Mg2+	372	0.1537	422	0.1278
1-Pb2+	418	0.1004	471	0.0037
1-Ca2+	525	0.1629	588	0.3722

In case of bikaverin (1), the HOMO and LUMO are spread evenly over the benzoquinone and hydroquinone rings of the molecule. The observed changes in the HOMO and LUMO contours of bikaverin (1) indicate a pi to pi* transition in absorption. However, HOMOs and LUMOs in the receptor-analyte adducts are budged on different regions of the adducts indicating intramoleculor charge transfer due to the binding of cationic species.

It can also be seen from the Fig. 3 that the orbital contour distribution of HOMO in case of bikaverin calcium ion adduct (1-Ca²⁺) gets shifted towards the opposite end compared to free bikaverin (1) or 1-Al³⁺ and 1-Mg²⁺ adducts. The calculated fluorescence wavelength for bikaverin (1) was found to be 576 nm (2.15 eV) with oscillator strength of 0.0102, depicting low intensity fluorescence peak in bikaverin (1). While the calculated fluorescence spectrum of 1-Ca²⁺ shows an intense peak at 588 nm (2.19 eV) with oscillator strength of 0.3722.

Thus the observations of strong Ca²⁺ ion binding coupled with the enhancement in the fluorescence intensity of bikaverin (1) on Ca²⁺ ion binding predict the bikaverin

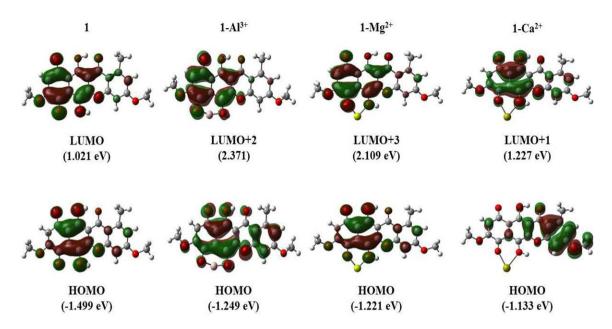


Fig. 3. Frontier molecular orbitals of bikaverin (1), 1-Al³⁺, 1-Mg²⁺ and 1-Ca²⁺.

(1) as the new possible fluorescent biosensor for Ca^{2+} ion. The fluorescence imaging of Ca^{2+} ions in the form of bikaverin (1) calcium adduct can serve as a method for analyzing signaling pathways involving calcium ions. Besides, the changes in the intracellular Ca^{2+} ion concentrations have

been related to different physiological, pathological and immune responses, therefore bikaverin based calcium ion sensing can also be an important tool in diagnostics.

The calculated oscillator strengths of fluorescence intensities of studied bioactive metal ions with bikaverin

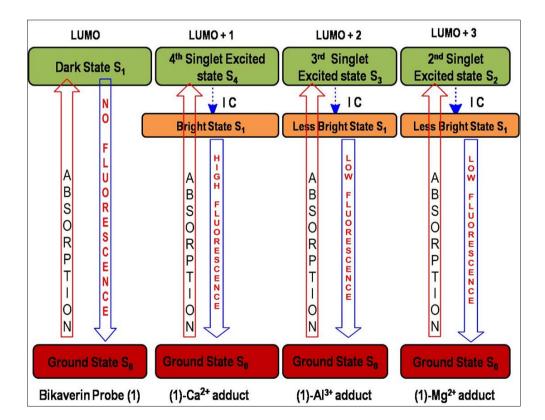


Fig. 4. Predicted electronic excitations and de excitations of bikaverin receptor (1) and its calcium adduct.

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(1) also predict a mild to medium increase in the intensity in case of 1-Mg²⁺ and 1-Al³⁺ adducts. But the intensities of their fluorescence peaks are predicted to be smaller in comparison to Ca²⁺ as depicted by their oscillator strengths of 0.1976 and 0.1278, respectively. For the other cationic species, there is no significant effect on the fluorescence intensities once they bind with the bikaverin receptor (1) as can be seen from their oscillator strengths in Table 3. It is therefore predicted from the DFT/TD-DFT calculations that receptor (1) can strongly sense Ca²⁺ ions fluorometrically with the milder sensing ability towards Mg²⁺ and Al³⁺ among the studied bioactive metal ions. The probable reason for the fluorescence intensity enhancement in case of 1-Ca²⁺, 1-Al³⁺, and 1-Mg²⁺ adducts is that their absorption transitions involve electron shifting to higher singlet excited states like fourth singlet excited state (S4), third singlet excited state(S3) and second singlet excited state(S2) in case of 1-Ca²⁺, 1-Al³⁺, and 1-Mg²⁺ respectively. As per Kasha's rule, 34 the fluorescence phenomenon occurs from the firstsinglet excited state (S1). In order for fluorescence to occur, the higher excited states have to relax to the first singlet excited state. For this relaxation process, internal conversions and the vibrational relaxations occur which lead to the loss of energy during the de-excitation. Due to this energy loss there is significant Stoke's shift (change between absorption and emission energies) While as in the other cations and in receptor (1), the transitions involve the lowest singlet excited state S1. So there is no internal conversion and hence no energy loss and eventually no Stoke's shift. Hence, in such cases, there is either no fluorescence or the intensity of the fluorescence is very weak.

TD-DFT calculated results show that the main transitions for bikaverin(1), 1-Al³⁺, 1-Mg²⁺ and 1-Ca²⁺ are

from HOMO \Rightarrow LUMO, HOMO \Rightarrow LUMO+2, HOMO \Rightarrow LUMO+3 and HOMO \Rightarrow LUMO+1 respectively see Fig 4. In all other receptor-analyte complexes, the main transitions involve HOMO \Rightarrow LUMO.

The experimental and theoretical fluorescence study of bikaverin (1) reveals major fluorescence peak corresponding to HOMO to LUMO transition with λ_{em} 576 nm in the theoretical spectra and λ_{em} 425 nm in experimental spectra. However on binding of Ca²+ the theoretical fluorescence peak gets shifted to a value λ_{em} 588 nm which closely corroborateswith the observed experimentalpeak of Bikaverin Ca²+adduct around λ_{em} of 620 nm.Thus, the theoretically predicted red shift in fluorescence of bikaverin(1) on calcium ion binding was also experimentally observed and therefore can be rationalized with theoretical prediction of HOMO \rightarrow LUMO+1 absorption in case bikaverin calcium ion adduct.

3. 1. Characterization of Bikaverin from Fungal Extract

The developments in synthetic biology has allowed tuning of microbial systems for generating industrially viable strains as the green and sustainable sources of value-added compounds.³⁵The composition and amounts of metabolites(bikaverin) are specific to the species of genus Fusarium. The morphological features of our crude fungal extract matched to the species *Fusarium proliferatum*. The chemical structure of purified bikaverin was confirmed through comparative analysis of NMR, mass, λmax and TLC of isolated compound with bikaverin standard and further to the literature reported values.³⁶ ¹H-NMR (CDCl3, 400MHZ):δ 2.87 (3H, s, 1-Me), 3.93

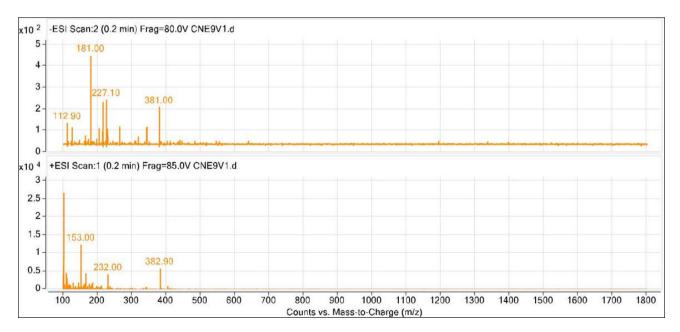


Fig. 5. Mass spectra of isolated product Bikaverin

(3H, s, 8-OMe), 3.96 (3H, s, 3-OMe), 6.35 (1H, s, H-9), 6.81 (1H, s, H-2), 6.93 (1H, s, H-4).HRMS (ESI⁺) calculated for C20H14O8 [M+H]⁺ : 381.0761, found: 382.90 Fig.5.

3. 2. Metal Ion Sensing Applications

The preliminary metal ion sensing behavior of bikaverin (1) was studied towards selected bioactive metal ions as their chloride/perchlorate salts in CH₃CN/H₂O (6:4, v/v; buffered with PBS, pH 7.4) using fluorescence spec-

troscopy Fig 6. The fluorescence spectrum of receptor (1) (10.0 μ M) exhibits a weak emission band at 620 nm corresponding to bikaverin moiety when excited at 596 nm in CH₃CN/H₂O (6:4, v/v; buffered with PBS, pH = 7.4). The weak fluorescence emission behavior of bikaverin is due to intramolecular charge transfer process as seen from HOMO- LUMO orbital contours.

It is evident from Figure 7 that on binding of calcium ions to bikaverin (1) its fluorescence profile gets modified. The free bikaverin shows very little to no fluorescence when excited at 596 nm while as the calcium bound bika-

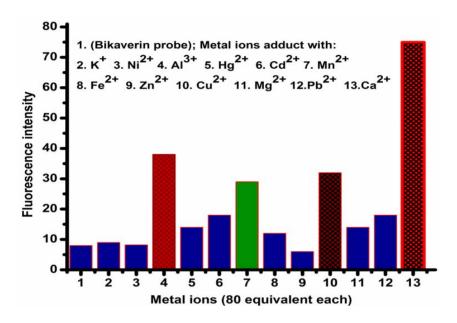
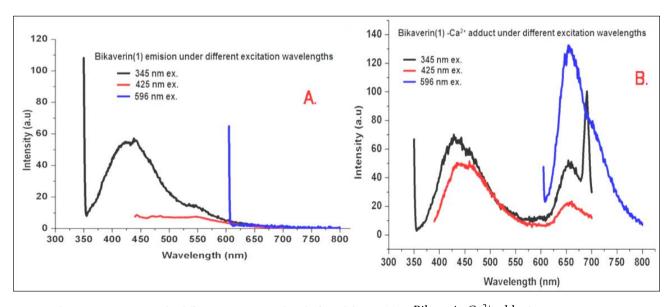


Fig. 6. Fluorescence response of bikaverin (1) towards various metal ions: Bars represent fluorescence selectivity (I/Io) of bikaverin (1) (10.0 μ M) towards various metal ions (80 equiv each) in CH3CN:H2O (6:4, v/v) buffered with 0.05 M HEPES, pH = 7.4; λ ex = 596 nm. The experimental fluorescence spectra of bikaverin(1) and its calcium adduct under different excitation wavelengths are shown in Fig.7 (A,B) respectively.



 $\textbf{Fig. 7}. \ \ \text{Fluorescence response under different excitation wavelengths for A: bikaverin (1) B: Bikaverin Ca^{2+} \ adduct.$

verin shows a major fluorescence when excited at the 596 nm wavelength. Among the fluorescence peaks obtained under the excitation wavelengths of 425 and 345 nm, the former gets an enhancement in the emission intensity while as the latter peak remains largely unchanged on calcium ion binding to bikaverin.

4. Conclusions

In summary, the present work describes a thorough computational investigation of metal ion sensing ability of natural pigment bikaverin as a fluorescent probe. An exploration of comparative metal ion binding affinities and optical properties of the bikaverin adducts through systematic studies using the appropriate level of theory are presented. The Ca²⁺, Mg²⁺and Al³⁺ metal ions are shown to bind the bikaverin receptor more strongly and with an enhancement in its florescence intensity. The strongest binding affinity with the most intense fluorescence emission in case of Ca²⁺ predicts bikaverin pigment as a possible Ca²⁺ biosensor. The concept of isolation of bikaverin receptor (1) from the natural source as an economical and sustainable method is also presented. The computationally predicted metal ion fluorescence behavior of bikaverin receptor (1) towards the studied metal ions was experimentally corroborated through the preliminary fluorescence studies. Among the studied metal ions, the bikaverin receptor (1) was seen to strongly detect Ca²⁺ions with an enhancement of fluorescence emission in its µM concentration range in the buffered aqueous medium due to charge transfer phenomena. The inclusive experimental studies of the bikaverin receptor (1) towards the selective calcium ion sensing under physiological conditions, the complete analytical profile of bikaverin as biosensor, the real time imaging of Ca²⁺ ions in cells through confocal microscopy and other related studies for the buildup of bikaverin as Ca²⁺ ion biosensor are underway in our laboratory.

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Conflict of Interest

All the authors declare no conflict of interest what so ever.

6. References

- H. Nikoofard, M. Sargolzaei, F. Faridbod Acta Chim. Slov. 2017, 64, 842–848. DOI:10.17344/acsi.2017.3357
- S. Krishnamurty, M. Stefanov, T. Mineva, S. Bégu, J. M. Devoisselle, A. Goursot, R. Zhu, D. R. Salahub, *J. Phys. Chem. B.* 2008, 112, 13433–13442. DOI:10.1021/jp804934d
- G. J. Cheng, X. Zhang, L. W. Chung, L. Xu, Y. D. Wu, J. Am. Chem. Soc. 2015, 137, 1706–1725. DOI:10.1021/ja5112749
- 4. F. Odame, *Acta Chim. Slov.* **2018**, *65*, 328–332 **DOI**:10.17344/acsi.2017.4001
- A. T. Castro, J. D. Figuero-Villar, *Int. J. Quantum Chem.* 2002, 89, 135–146. DOI:10.1002/qua.10302
- M. Lintuluoto, J. M. Lintuluoto, *Biochemistry*. 2016, 55, 4697–4707. DOI:10.1021/acs.biochem.6b00423
- S. Arulmozhiraja, M. Morita, Chem. Res. Toxicol. 2004, 17, 348–356. DOI:10.1021/tx0300380
- 8. N. Yasarawan, K. Thipyapong, V. Ruangpornvisuti, *J. Mol. Graphics Model.* **2014**, *51*,13-26. **DOI**:10.1016/j.jmgm.2014.04.009
- F. Senn, M. Krykunov, J. Phys. Chem. A. 2015, 119, 10575– 10581. DOI:10.1021/acs.jpca.5b07075
- M. H. Bridge, E. Williams, M. E. G. Lyons, K. F. Tipton, W. Linert. *Biochim. Biophys. Acta.* 2004, 1690, 77–84.
 DOI:10.1016/j.bbadis.2004.05.007
- Y. Wu, B. Midinov, R.J. White, ACS Sens., 2019, 4, 498–503.
 DOI:10.1021/acssensors.8b01573
- M. A. Rizvi, M. Mane, M. A. Khuroo, G. M. Peerzada, Monatsh Chem. 2017, 148, 655–668.
 DOI:10.1007/s00706-016-1813-8
- Y. Dangat, M. A. Rizvi, P. Pandey, K. Vanka, *J Organomet Chem.* 2016, 801, 30-41.
 DOI:10.1016/j.jorganchem.2015.10.015
- 14. M.V. Mane, M. A. Rizvi, K. Vanka, *J. Org. Chem.* **2015**, *80*, 2081–2091. **DOI**:10.1021/jo5023052
- M. Kumar, A. Kumar, M. Rizvi, M. Mane, K. Vanka, S. C. Taneja, B. A. Shah, *Eur. J. Org. Chem.* 2014, 2014, 5247–5255.
 DOI:10.1002/ejoc.201402551
- A. Pandey, M. Rizvi, B. A. Shah, S, Bani, Cytokine, 2016, 79, 103-113. DOI:10.1016/j.cyto.2016.01.004
- A. Goswami, B. A. Shah, A. Kumar, M. A. Rizvi, S. Kumar, S. Bhushan, F. A. Malik, N. Batra, A. Joshi, J. Singh, Chem.-Biol. Interact., 2014, 222, 60–67. DOI:10.1016/j.cbi.2014.08.011
- R Chib, M. Kumar, M. Rizvi, S. Sharma, A. Pandey, S. Bani,
 S Andotra, S. C. Taneja, B. A Shah, RSC Advances, 2014, 4,
 8632–8637. DOI:10.1039/c3ra46412a
- J. Balan, J. Fuska, I. Kuhr, V. Kuhrova, Folia Microbiol (Praha).
 1970, 15, 479–484. DOI:10.1007/BF02880192
- J. F. Henderson, M. L. Battell, G. Zombor, J. Fuska, P. Nemec, Biochem Pharmacol. 1977 26, 1973–1977.
 DOI:10.1016/0006-2952(77)90004-1
- D. Kjaer, A. Kjaer, C. Pedersen, J. D. Bulock, J. R. Smith, J. Chem Soc Perkin 1. 1971, 16, 2792–2797.
 DOI:10.1039/I39710002792
- 22. M. C. Limon, R. Rodríguez, J. Avalos, *Appl Microbiol Biotech-nol.* **2010**, *87*, 21–29. **DOI**:10.1007/s00253-010-2551-1

- D. Arora, N. Sharma, V. Singamaneni, V. Sharma, M. Kushwaha, V. Abrol, S. Guru, S. Sharma, A. P. Gupta, S. Bhushan, S. Jaglan, P. Gupta, *Phytomedicine*. 2016, 23, 1312–1320.
 DOI:10.1016/j.phymed.2016.07.004
- 24. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; et al. Gaussian 09; Gaussian Inc: Wallingford, CT, 2009.
- 25. T. Yanai, D. P. Tew, N. C. Handy, Chemical Physics Letters, 2004, 393, 51–57. **DOI:**10.1016/j.cplett.2004.06.011
- Jacquemin, D.; Planchat, A.; Adamo, C.; Mennucci, B. J. Chem. Theory Comput. 2012, 8, 2359–2372.
 DOI:10.1021/ct300326f
- H. R. Bhat, P. C. Jha, Chem. Phys. Lett. 2017, 669, 9–16.
 DOI:10.1016/j.cplett.2016.12.025
- 28. H. R. Bhat, P. C. Jha, *Phys. Chem. Chem. Phys.* **2017**, *19*, 14811–14820. **DOI**:10.1039/C7CP02287E
- H. R. Bhat, P. C. Jha, J. Phys. Chem. A. 2017, 121, 3757–3767.
 DOI:10.1021/acs.jpca.7b00502

- 30. S. F. Boys, F. Bernardi, *Mol. Phys.***1970**, *19*, 553–566. **DOI:**10.1080/00268977000101561
- 31. Reed, A. E.; Curtiss, L.; Weinhold, F. *Chem. Rev.* 1988, 88, 899–926. **DOI**:10.1021/cr00088a005
- J. W. Cornforth, G. Ryback, P. M. Robinson, D. Park, *J Chem Soc Perkin* 1971, 1.16, 2786-2788.
 DOI:10.1039/j39710002786
- R. M. Jagtap, M. A. Rizvi, Y. B. Dangat, Satish K. Pardeshi, *J. Sulfur Chem.* 2016, 37, 401–425.
 DOI:10.1080/17415993.2016.1156116
- 34. M. Kasha, *Discuss. Faraday Soc.* **1950**, *9*, 14–19. **DOI:**10.1039/df9500900014
- 35. J. Du, Z. Shao, H. Zhao, *J Ind Microbiol Biotechnol.* **2011,** 38, 873–890. **DOI:**10.1007/s10295-011-0970-3
- Kjaer, A. Kjaer, C. Pedersen, J. D. BuLock, J. R. Smith, J. Chem. Soc. C 1971, 0, 2792–ww2797.
 DOI:10.1039/J39710002792

Povzetek

Članek predstavlja računalniško raziskovanje gliv, ki proizvajajo pigment bikaverin kot biosensor za biološko razpoložljive kovinske ione. Narejene so bile sistematične študije geometrije optimiziranega osnovnega in vzbujenega stanja za raziskovanje vezavnega žepa za kovinske ione, primerjalna vezavna nagnjenost in optične lastnosti bikaverina in njegovih aduktov s preiskovanimi kovinskimi ioni. Pregled trinajstih (13) biodostopnih kovinskih ionov je pokazal, da jakost vezanja na bikaverinski receptor ustreza zaporedju Ca^{2+} , Mg^{2+} in Al^{3+} . Po vezavi na bikaverinski receptor je bila opažena ojačitev fluorescence v vrstnem redu $Ca^{2+} > Al^{3+} > Mg^{2+}$. Računsko napovedana selektivnost bikaverinskega receptorja za Ca^{2+} je bila eksperimentalno podprta s predhodnimi fluorescenčnimi študijami. Bikaverinska sonda je pokazala povečanje fluorescenčne emisije v prisotnosti Ca^{2+} ionov v zapufranem vodnem mediju.